



2022 MELISSA CONFERENCE
8-9-10 NOVEMBER 2022

CREATING
A CIRCULAR
FUTURE

Electroactive biofilm development under controlled hydrodynamic in a Couette-Taylor electrochemical reactor

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PROSe
PRocédés biOtechnologiques
au Service de l'Environnement


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ANR-17-CE06-0015-01, 2018-2023



Wastewater energetic impact : greywater

Tank oxygenation

75% for the plant



France : 3%

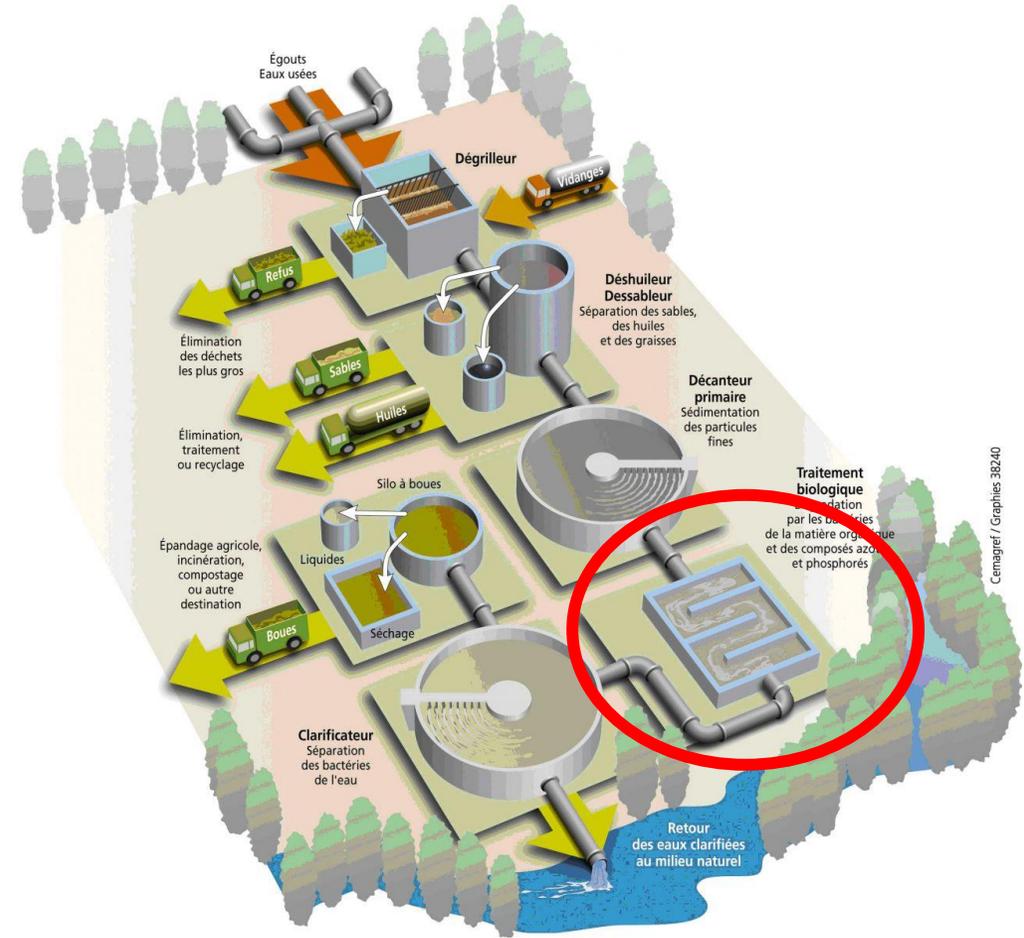


Figure 1 : Wastewater treatment process



Wastewater energetic impact : greywater

Tank oxygenation

75% for the plant



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Need of a way to decrease this consumption

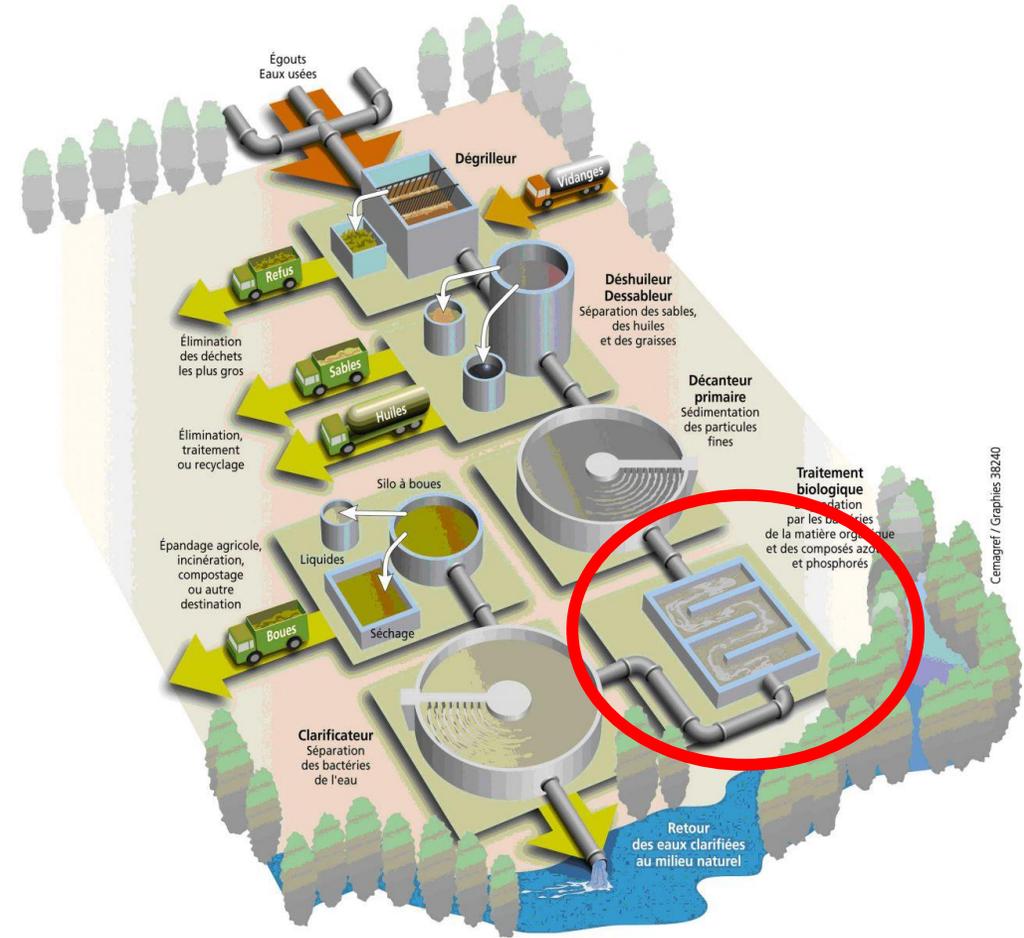


Figure 1 : Wastewater treatment process

Bio-Electrochemical Systems to separate the redox process in 2 compartments

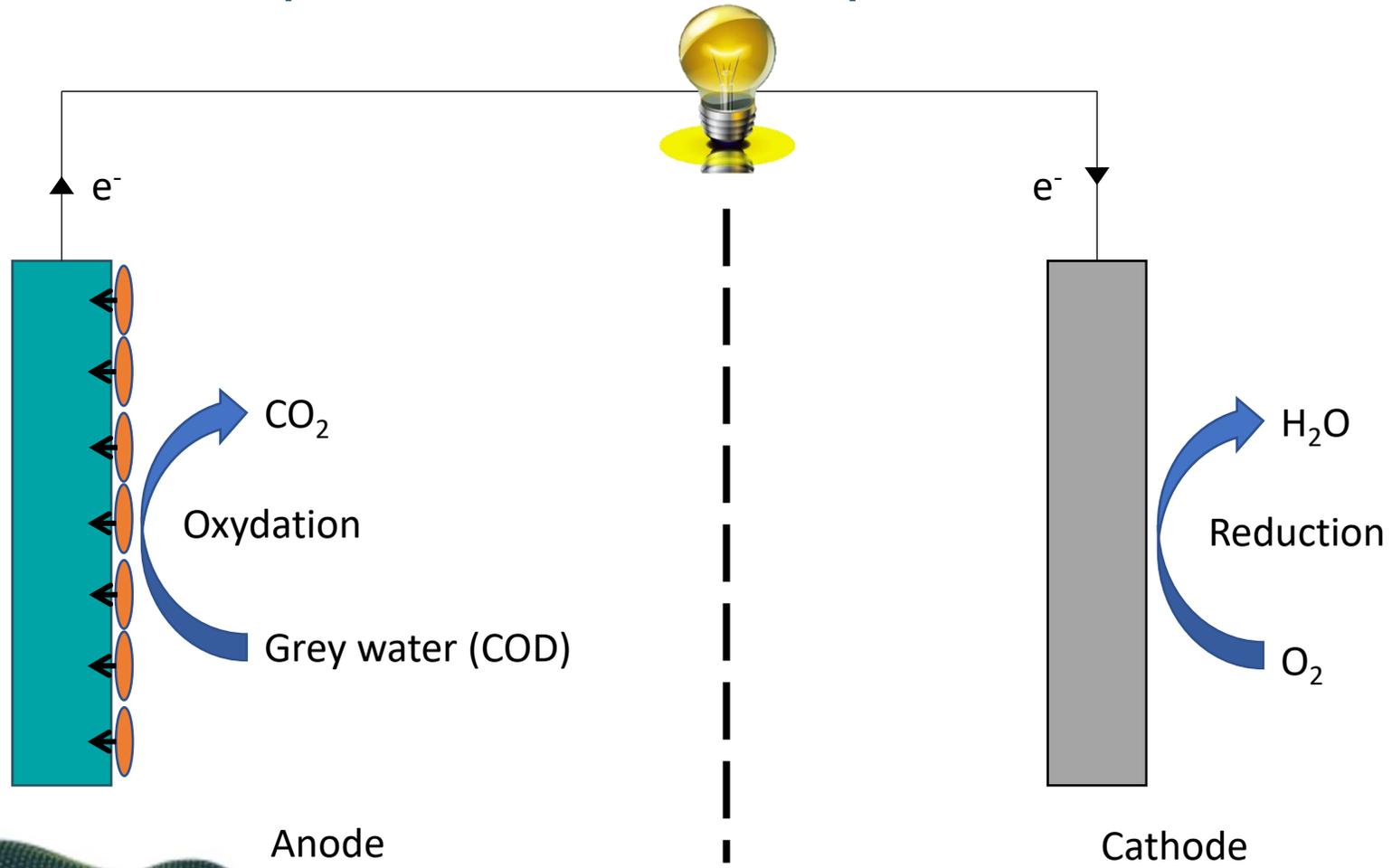
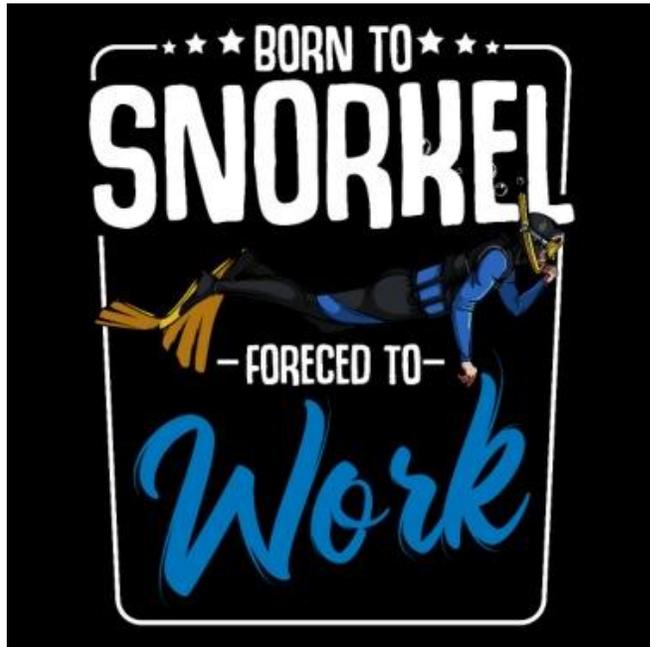


Figure 2 : The MFC principle



A bio-electrochemical snorkel : BIOTUBA





A bio-electrochemical snorkel : BIOTUBA

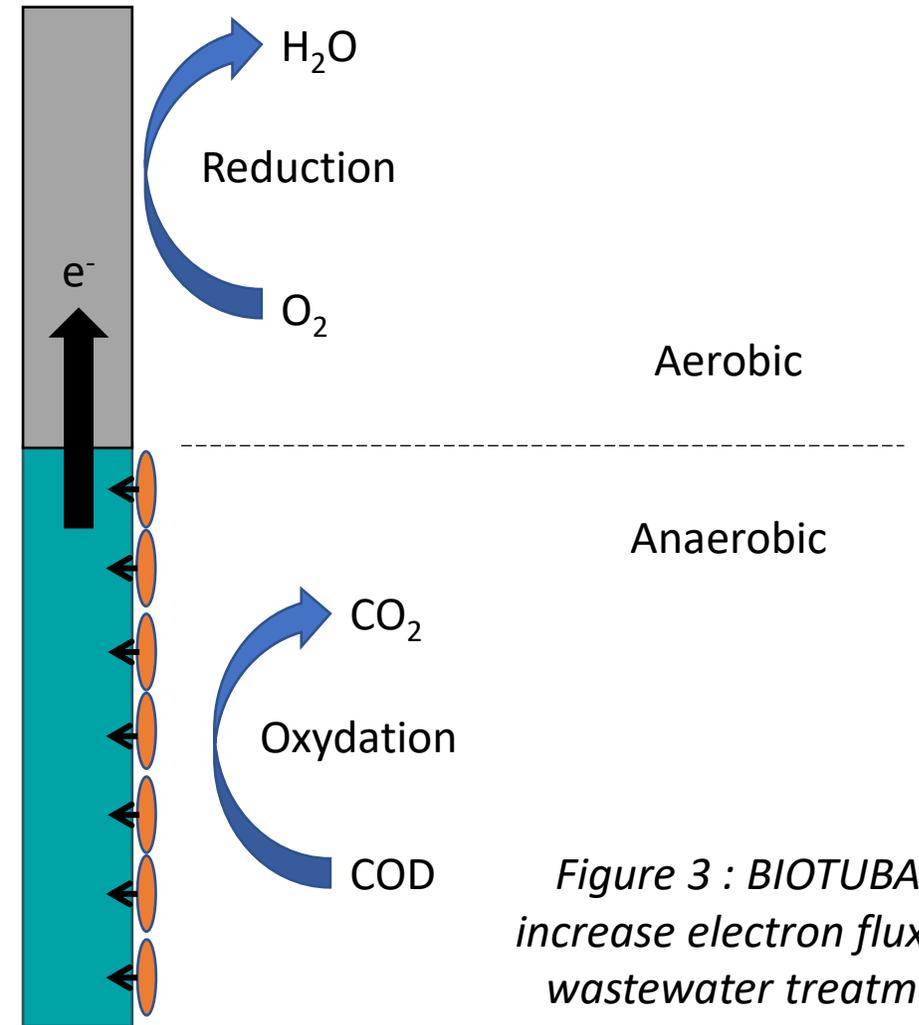
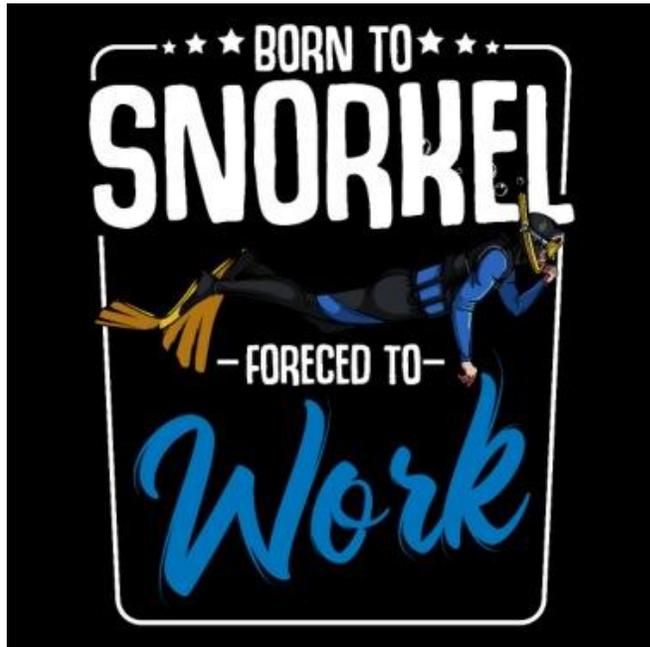
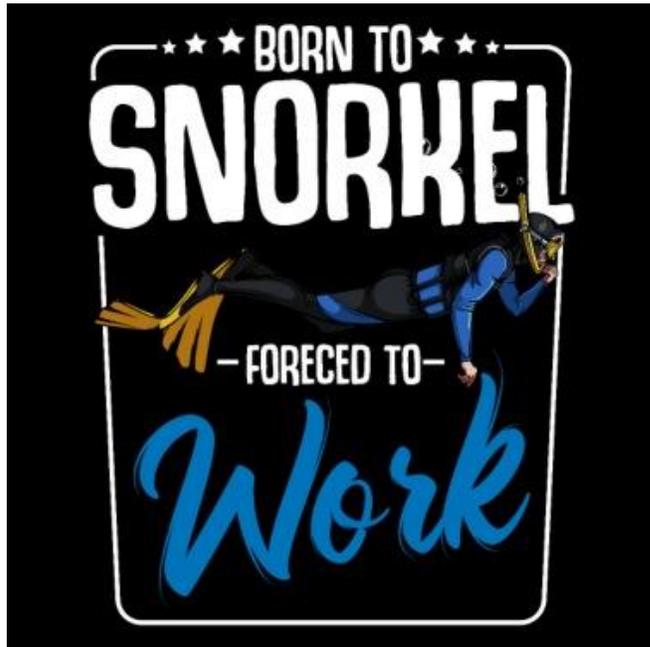


Figure 3 : BIOTUBA to increase electron flux and wastewater treatment efficiency



A bio-electrochemical snorkel : BIOTUBA



Limiting parameters

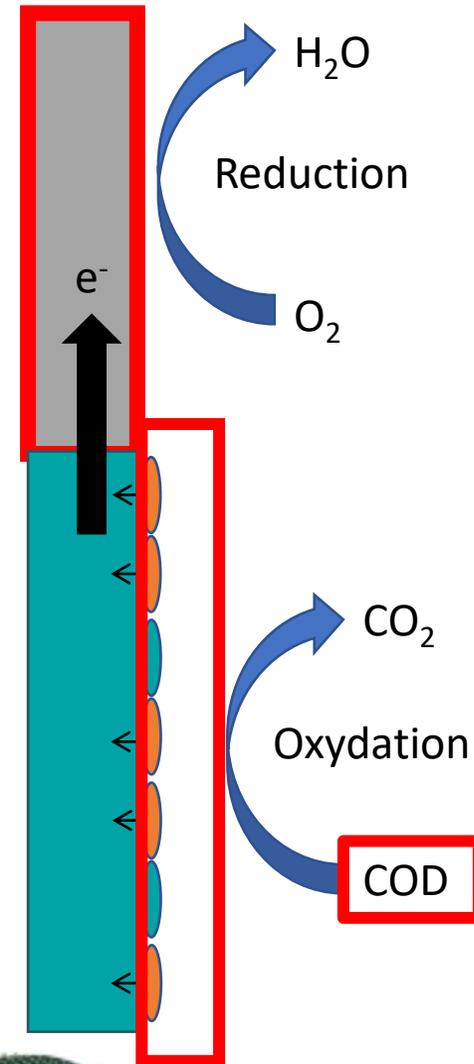
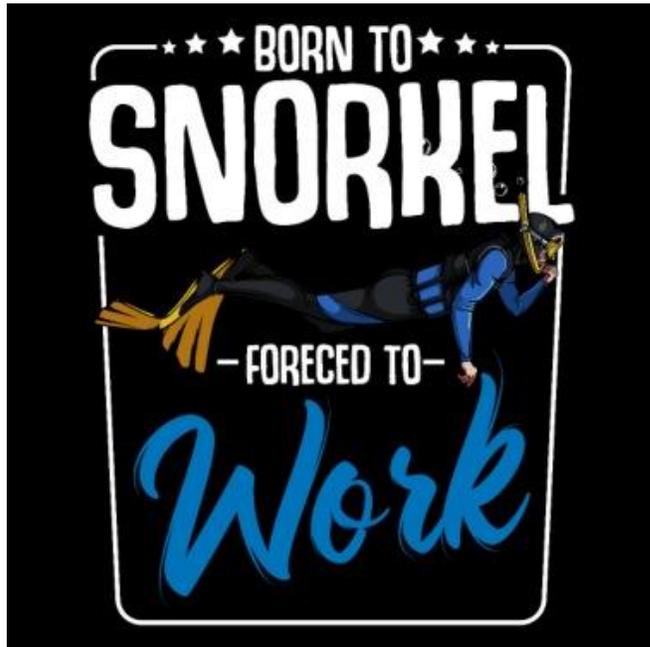


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A bio-electrochemical snorkel : BIOTUBA



Limiting parameters

Biofilm aging :
↳ Catalytic activity

➔ A way to boost the decreasing activity of aged biofilm?

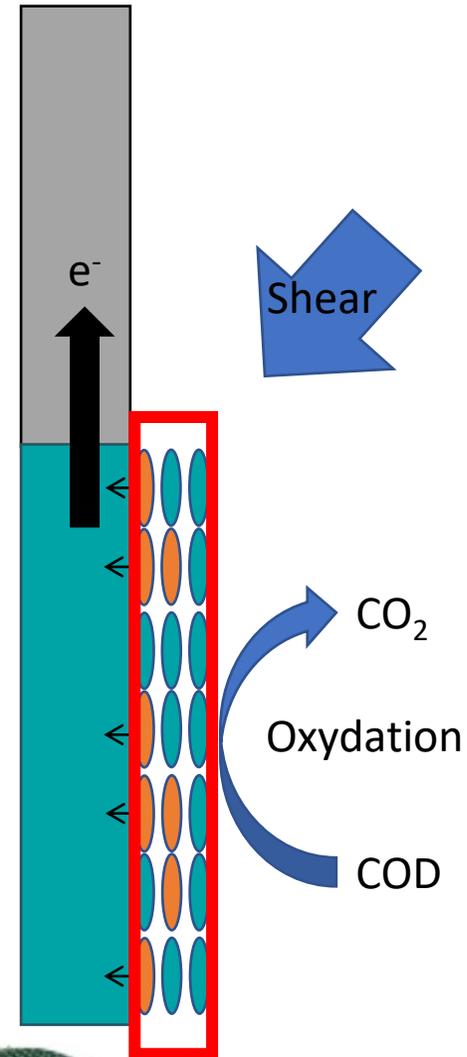
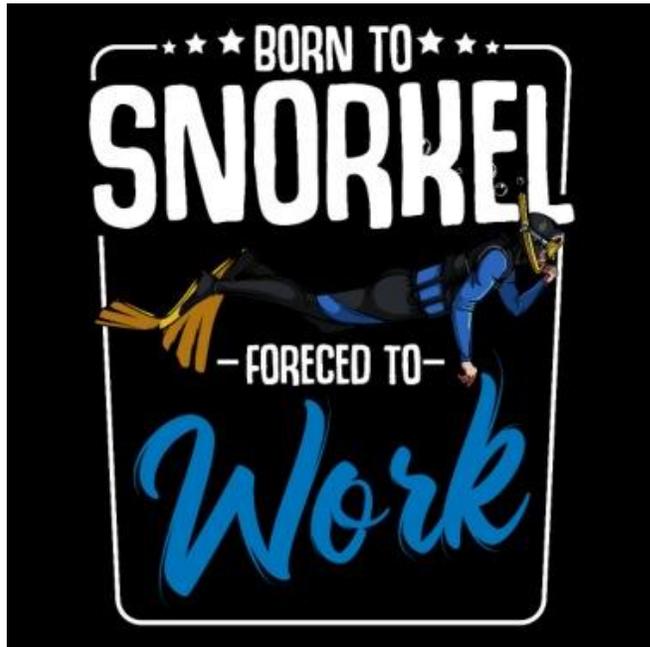


Figure 3 : BIOTUBA to increase electron flux and wastewater treatment efficiency

A bio-electrochemical snorkel : BIOTUBA



Limiting parameters

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➔ A way to boost the decreasing activity of aged biofilm?

?

Detachment of non-electroactive bacteria on the biofilm surface

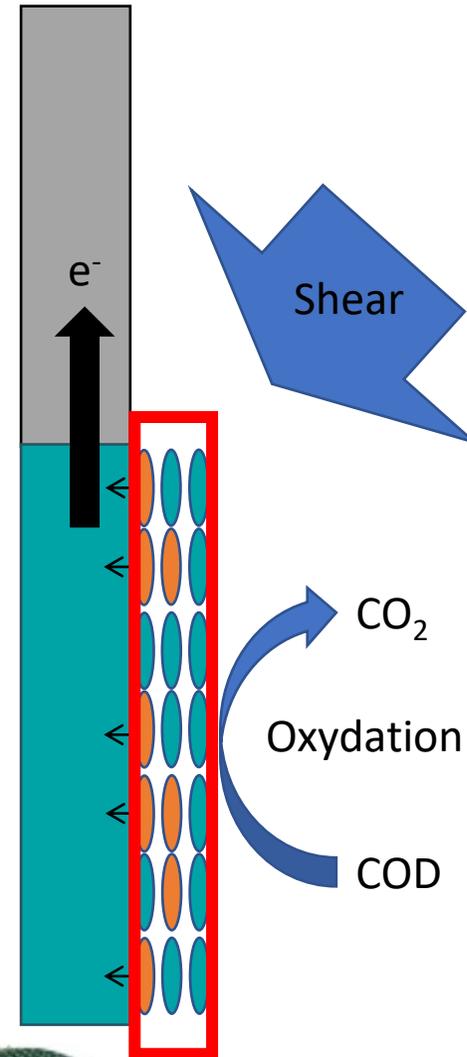
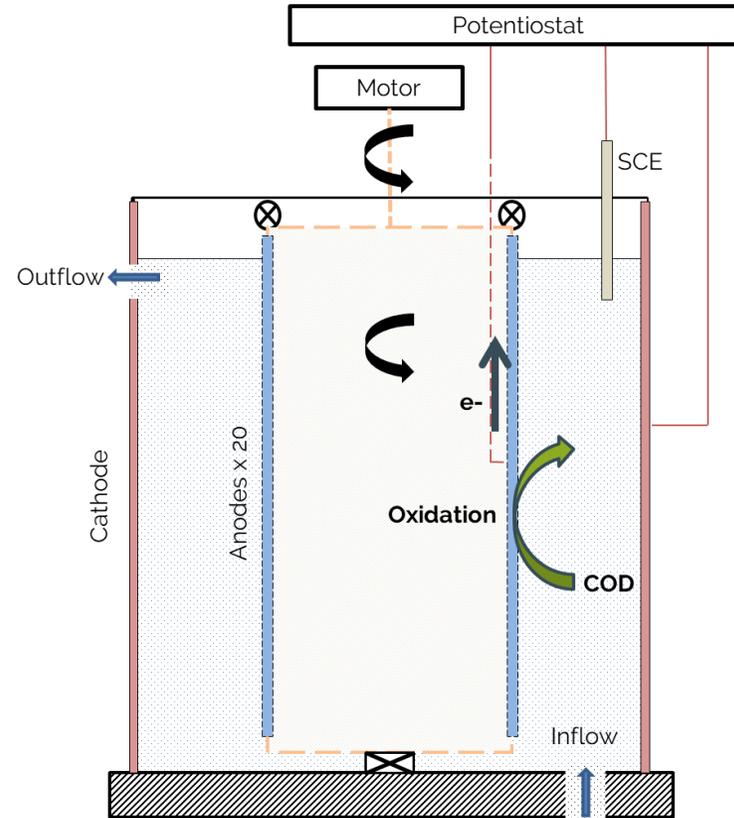


Figure 3 : BIOTUBA to increase electron flux and wastewater treatment efficiency

The first eCTR to characterize precisely the shear stress effect on electroactive biofilms



Advantages :

- **Decoupling** shear stress and other parameters (input flow rate or imposed voltage)

Figure 4 : Taylor-Couette reactor adapted to electroactive biofilm analysis

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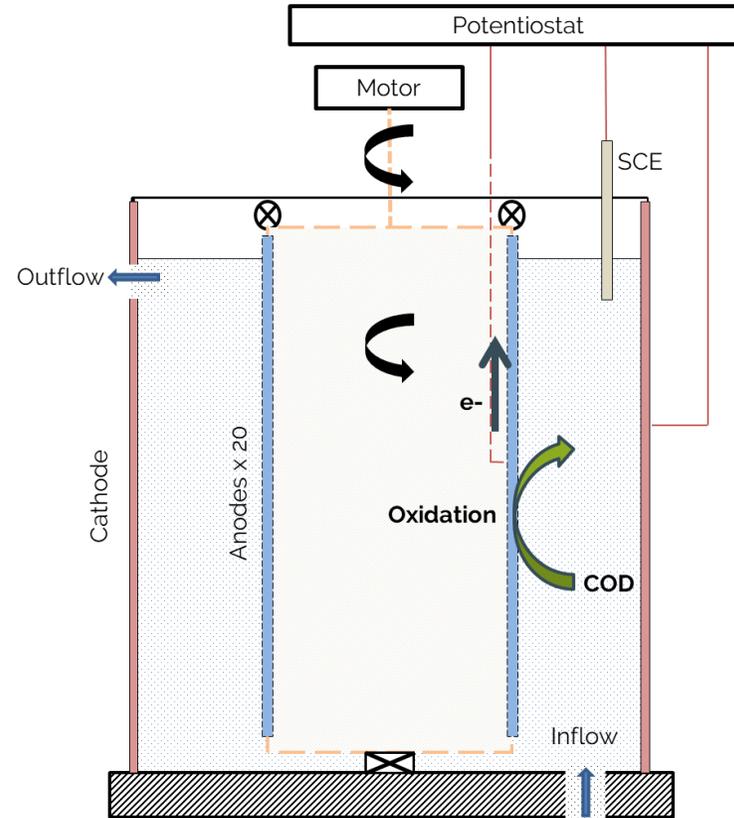


Figure 4 : Taylor-Couette reactor adapted to electroactive biofilm analysis

Advantages :

- **Decoupling** shear stress and other parameters (input flow rate or imposed voltage)
- Big size electrodes (20 cm²)
- Hydrodynamics **perfectly identical** for the 20 electrodes



Initiating the development of an electroactive biofilm under controlled low hydrodynamic

Reminder of the limiting parameters:

- Cathode size
- Substrate concentration
- Biofilm catalysis



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Operating conditions solutions:

- ✓ Studying different imposed voltage
- ✓ Real greywater supplemented with acetate
- ? Electrochemical (CA+CV) and biological (sequencing) analysis



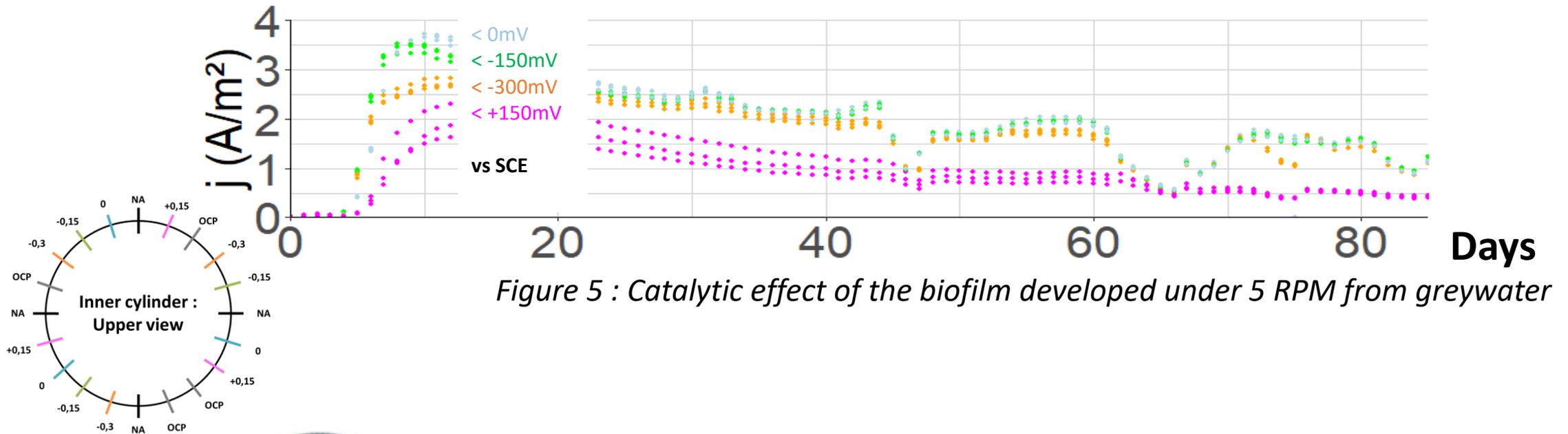
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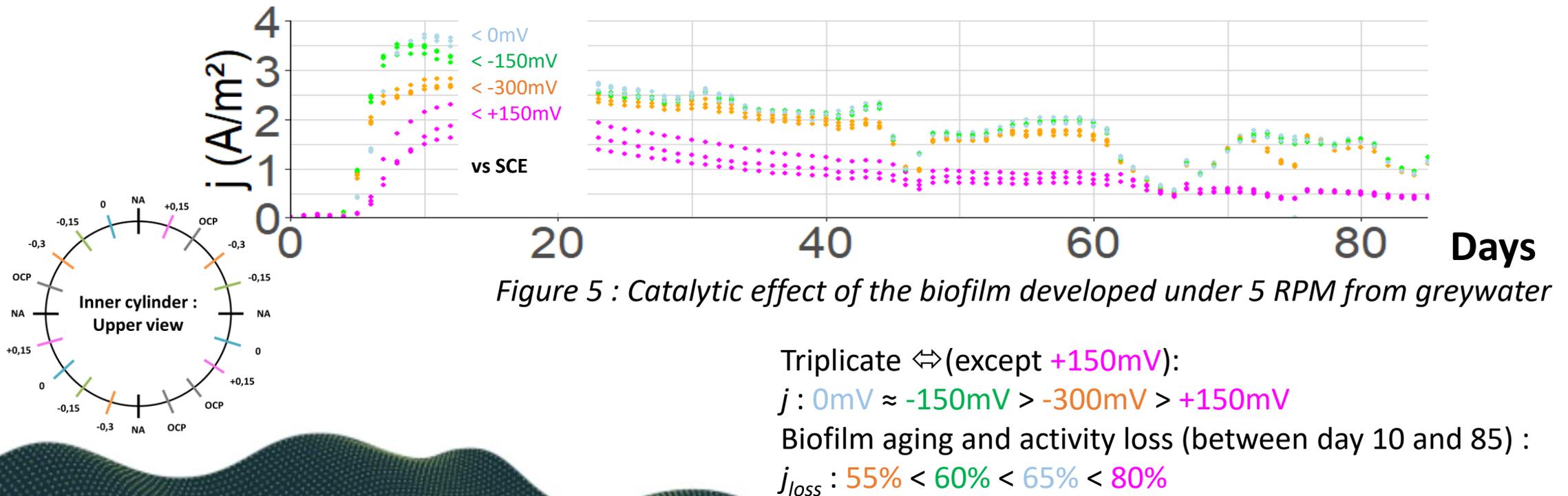




Initiating the development of an electroactive biofilm under controlled low hydrodynamic

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Characterizing electro-microbial anodic activity: Cyclic Voltammetry (CV)

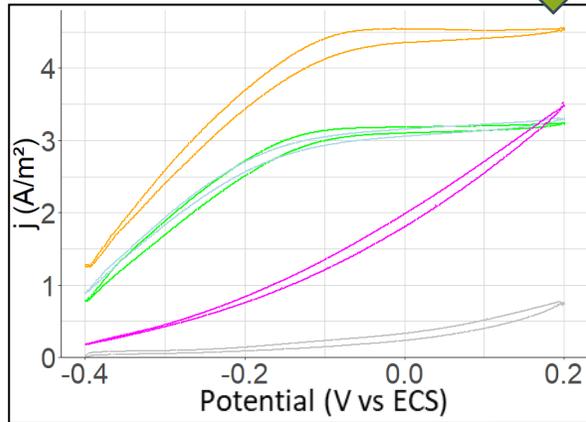
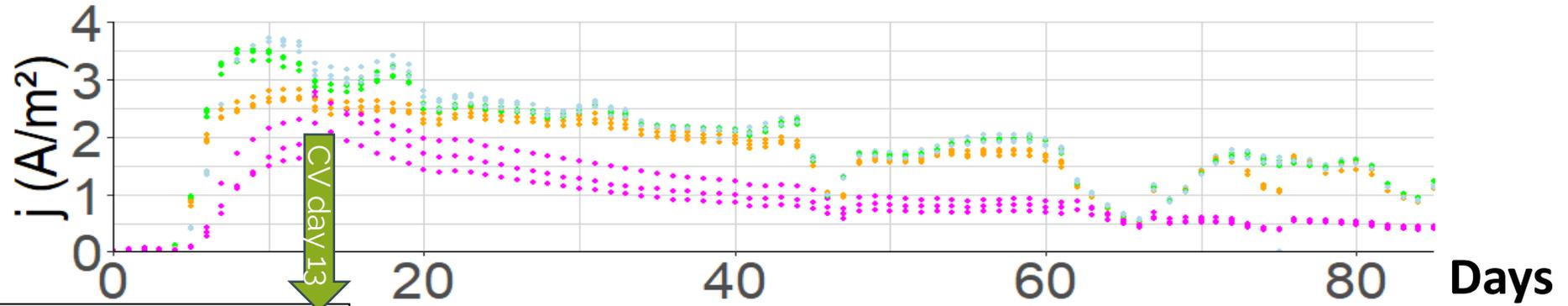


Figure 7 : CV
obtained at the
end of the
biofilm growth
phase

-300mV > -150mV = 0mV > +150mV

Lower catalysis effect



Characterizing electro-microbial anodic activity: Cyclic Voltammetry (CV)

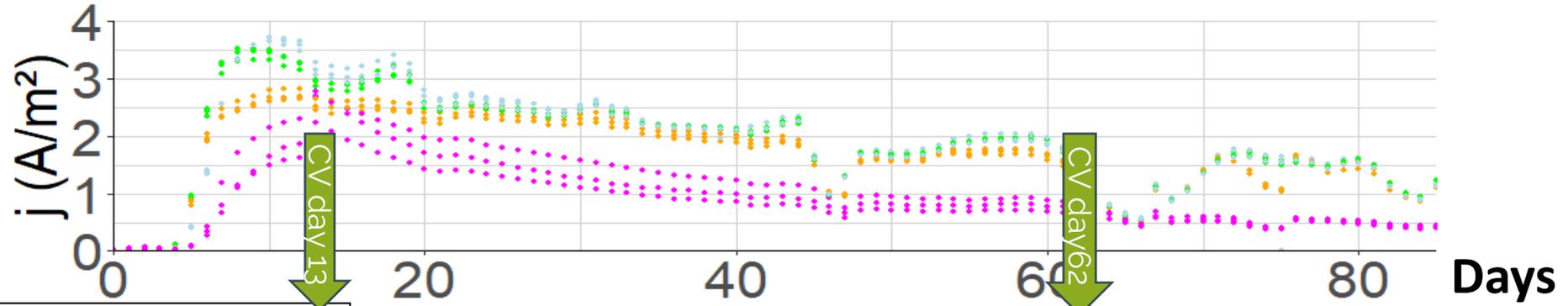
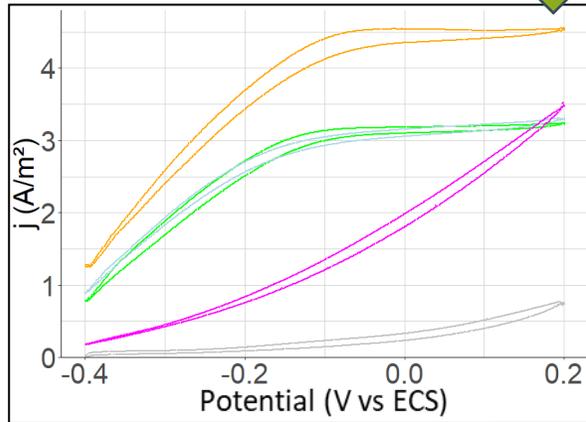


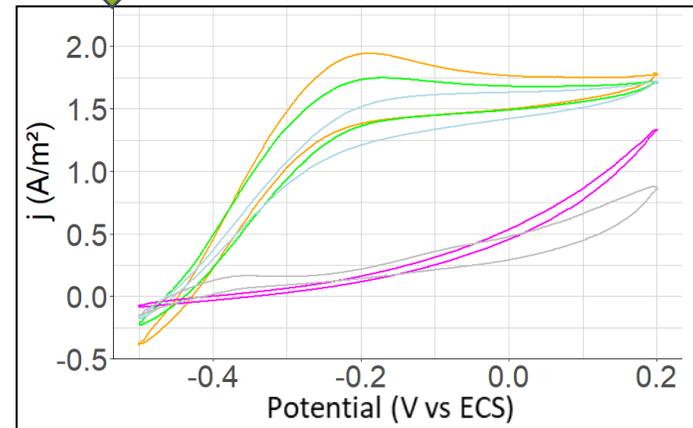
Figure 7 : CV obtained at the end of the biofilm growth phase



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Lower catalysis effect

Figure 8 : CV for the activity plateau of the biofilm



-300mV = -150mV = 0mV > +150mV

Converge

≈ 0



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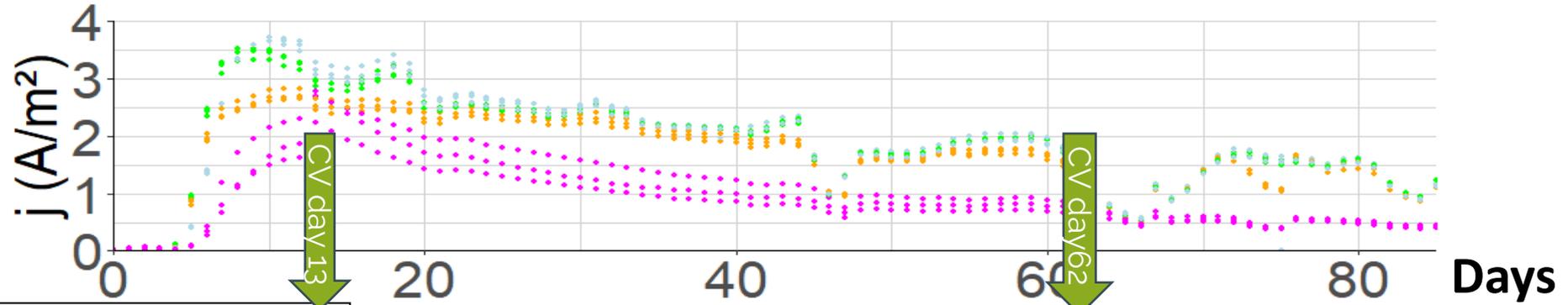
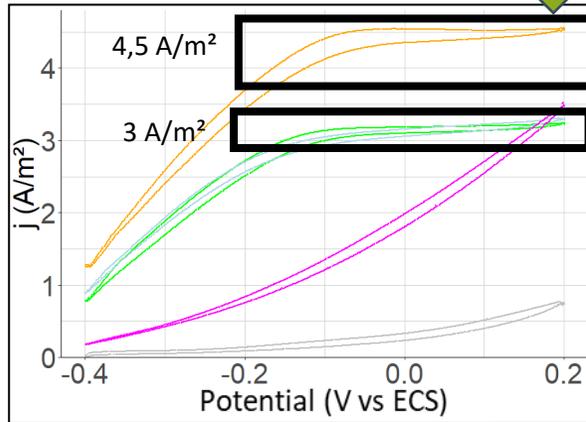


Figure 7 : CV obtained at the end of the biofilm growth phase



-300mV > -150mV = 0mV > +150mV

Lower catalysis effect

← Growth vs mature →

↘ j_{max} :

-60% / -50% / -50% / -65%

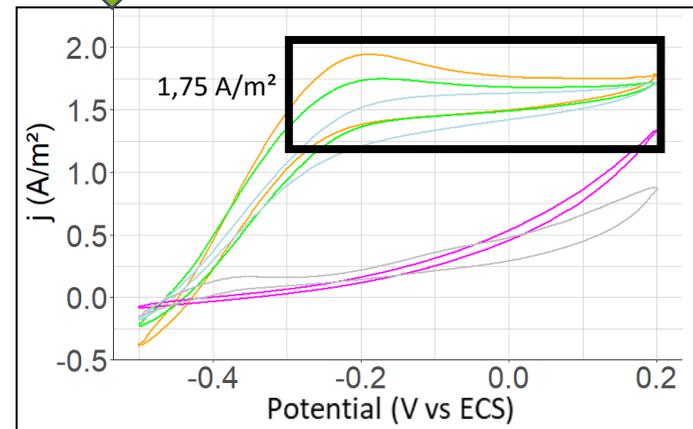


Figure 8 : CV for the activity plateau of the biofilm

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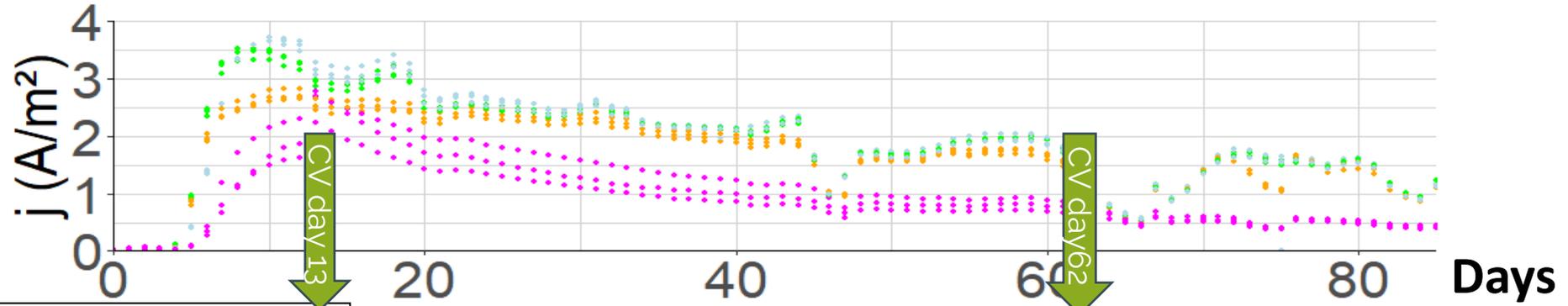
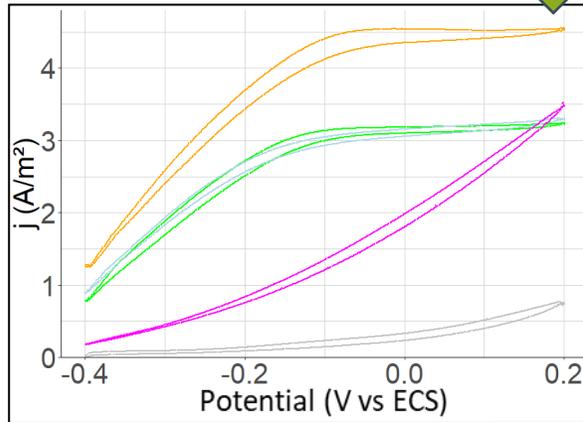


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Lower catalysis effect

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↗ Diffusion limitation phenomena

-300mV / -150mV

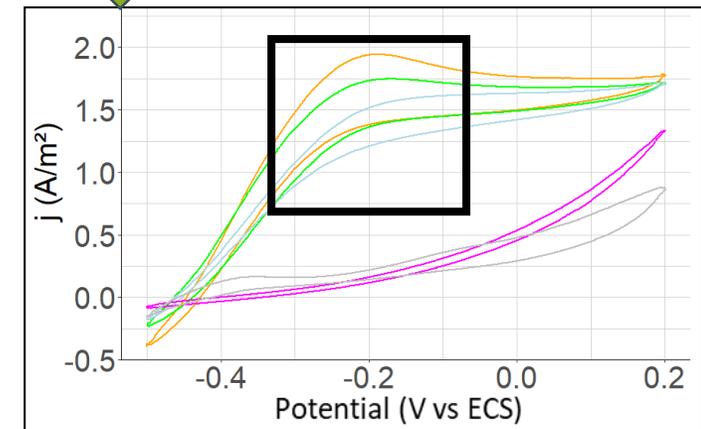


Figure 8 : CV for the activity plateau of the biofilm

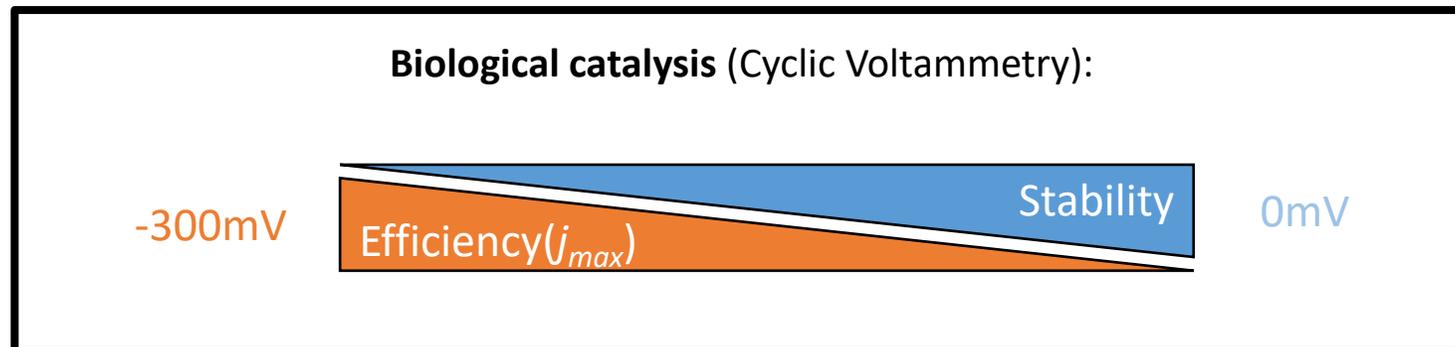
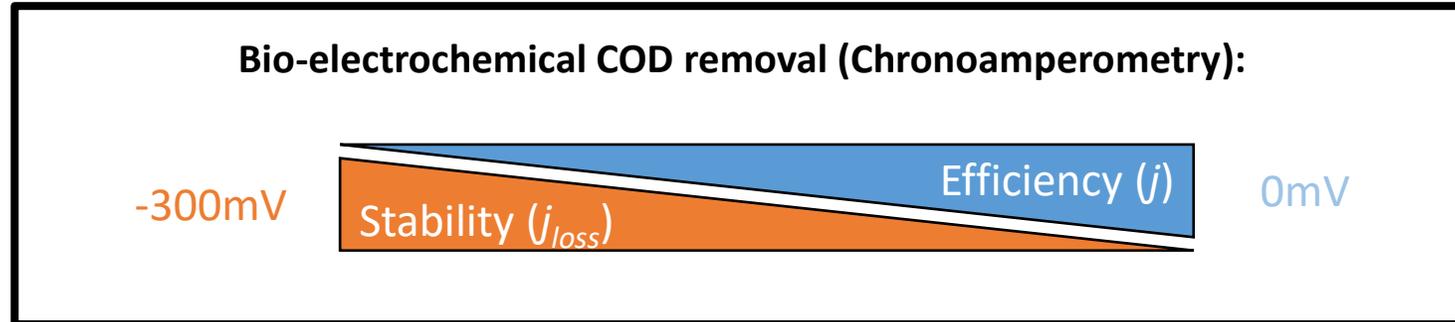
-300mV = -150mV = 0mV > +150mV

Converge

≈ 0



Electrochemical analysis summary



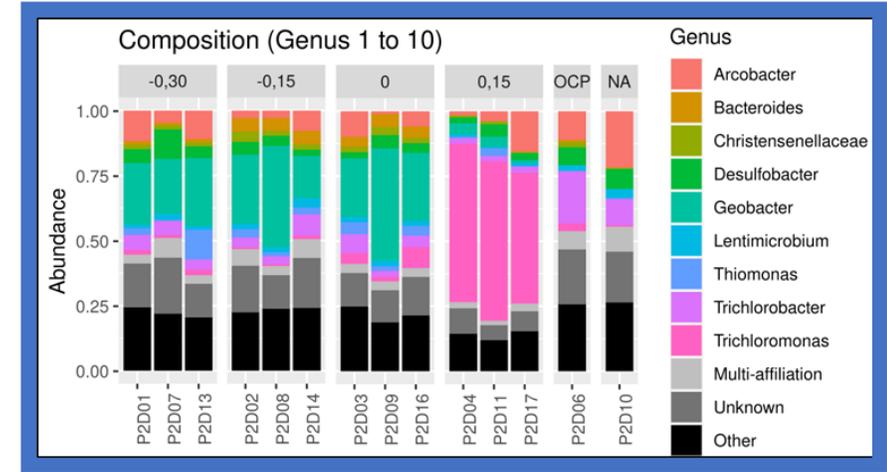
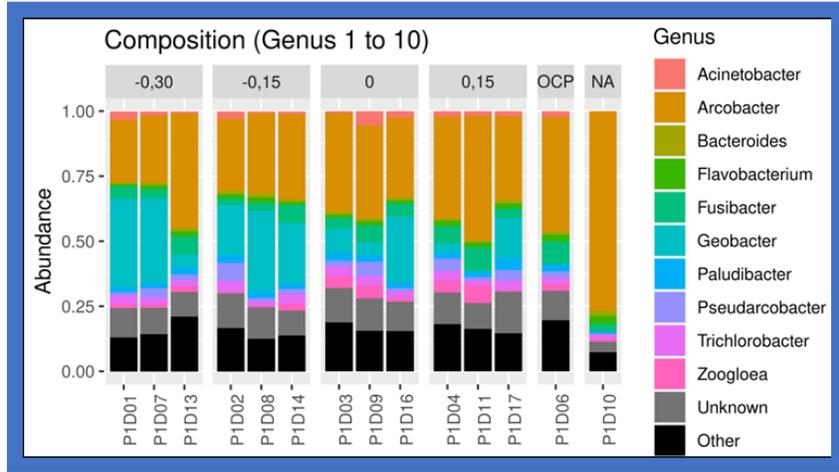


Microbial communities in the biofilm

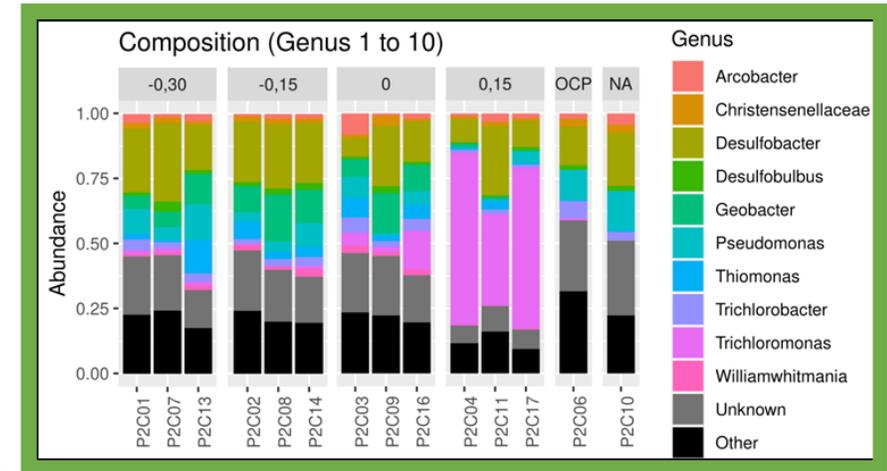
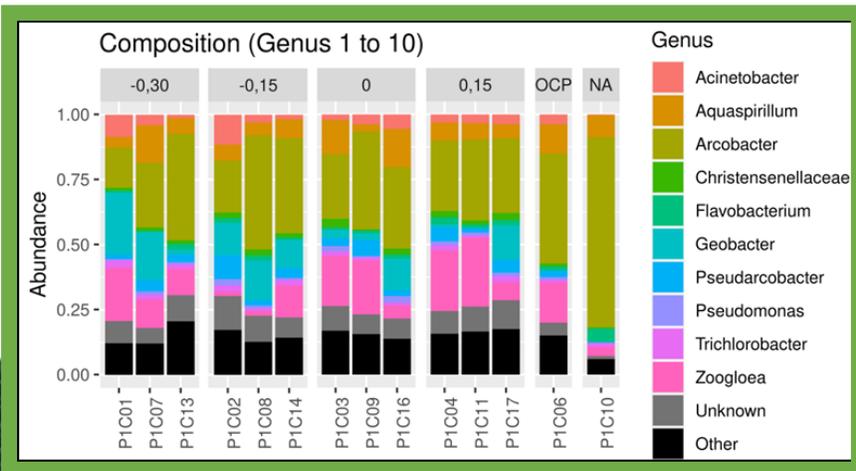
Day13

Day62

gDNA



cDNA

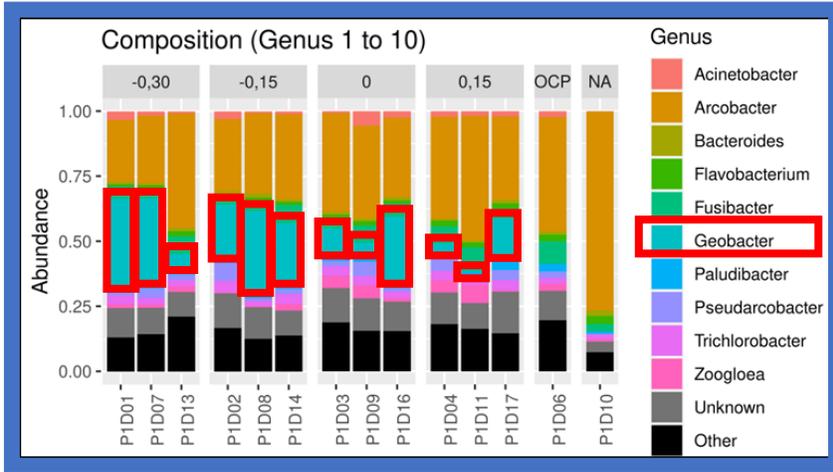




Day13

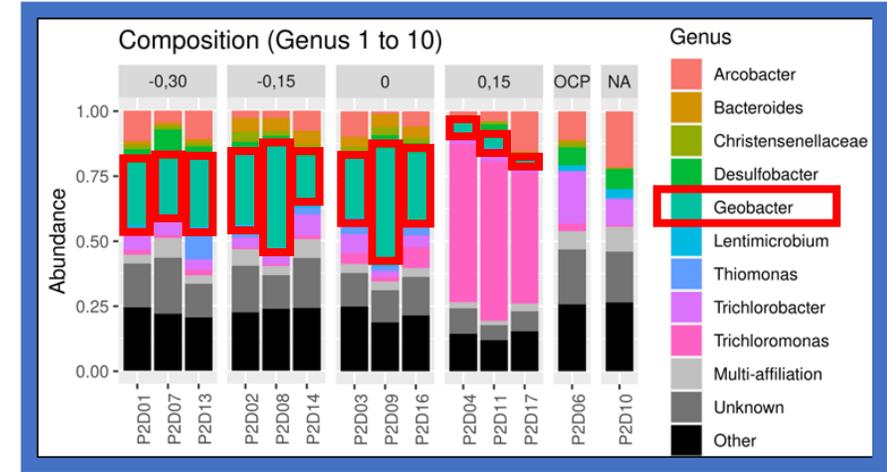
Geobacter, the electroactive bacterium

gDNA



In the most electroactive conditions:
 -300mV / -150mV / 0mV
 (↗: 20 -> 25%)

Day62



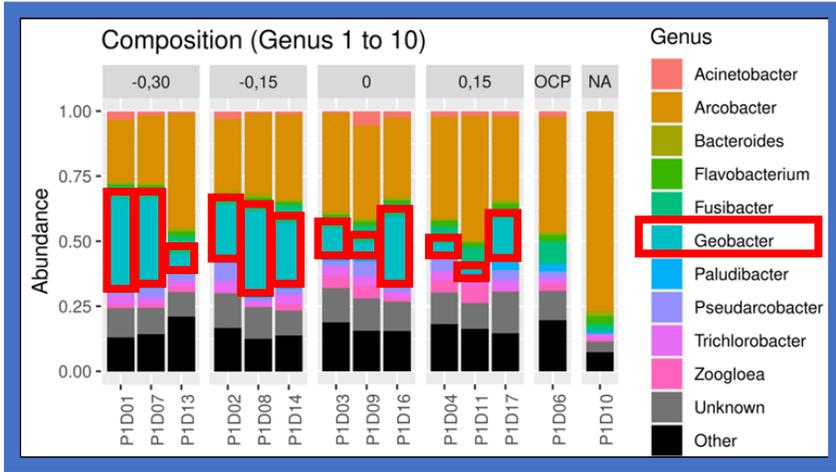


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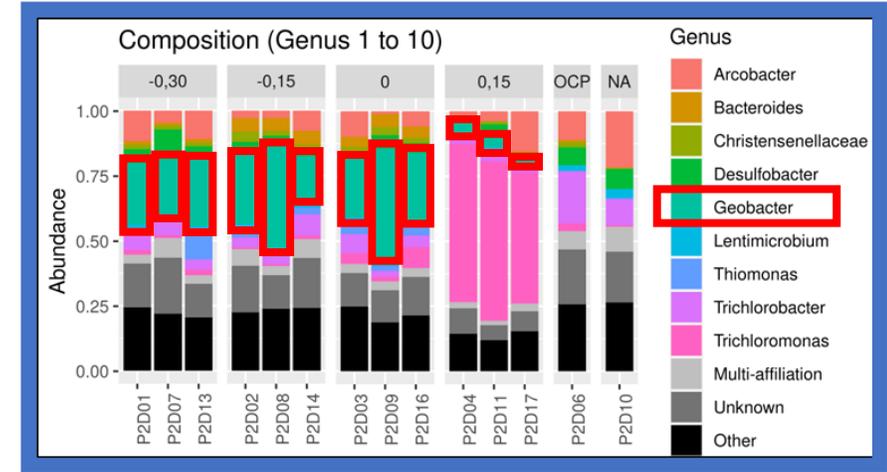
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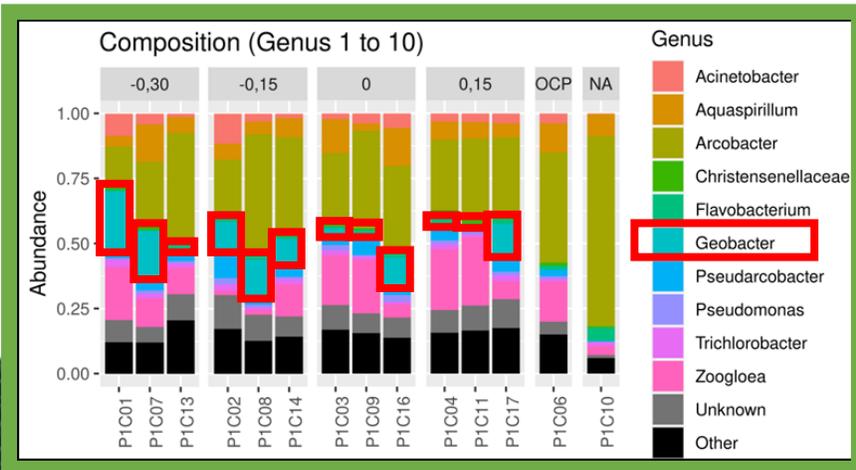
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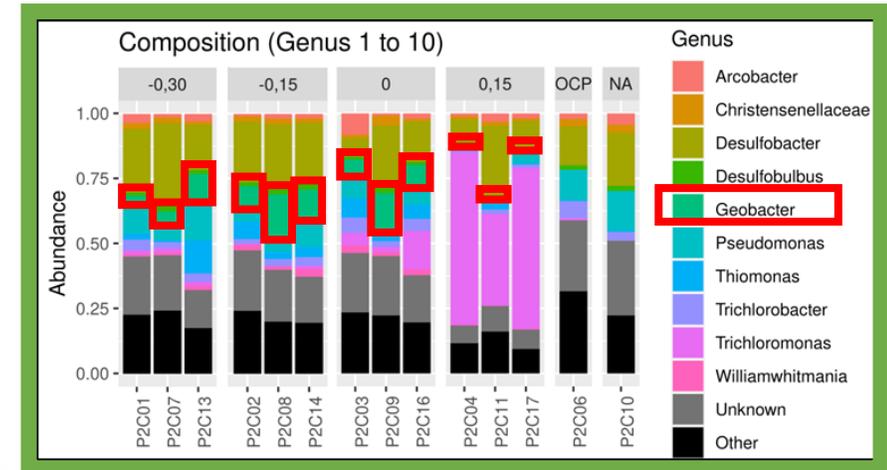
In the most electroactive conditions:
 -300mV / -150mV / 0mV
 (↗: 20 -> 25%)



cDNA



☐ Presence ++
 ☐ Activity - (10% -> 9%)

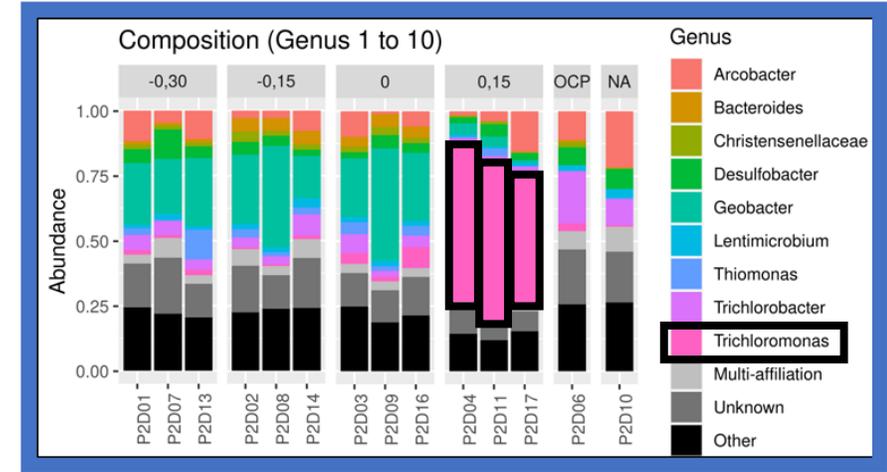
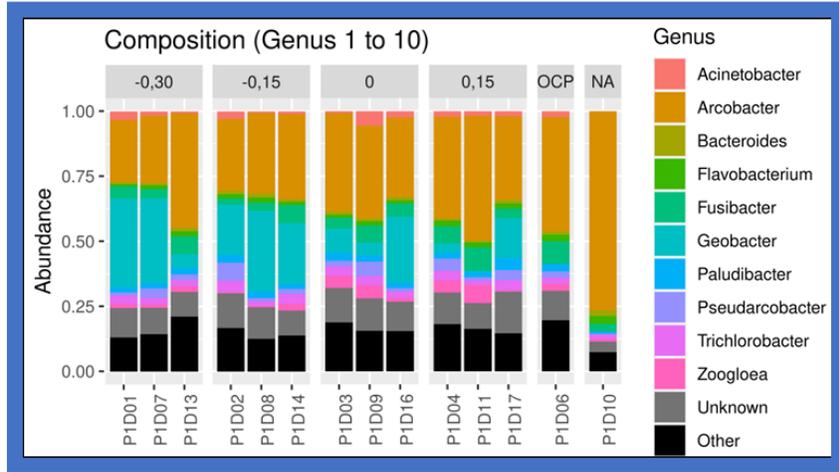


Hypothesis for *Trichloromonas*

Day13

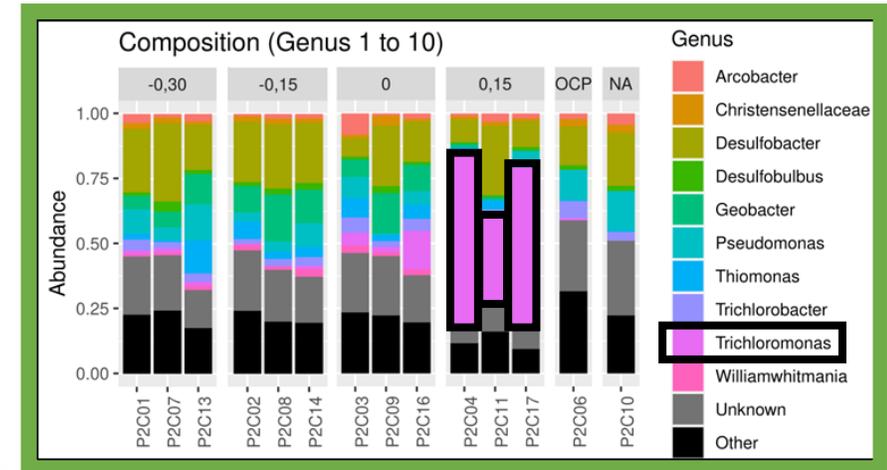
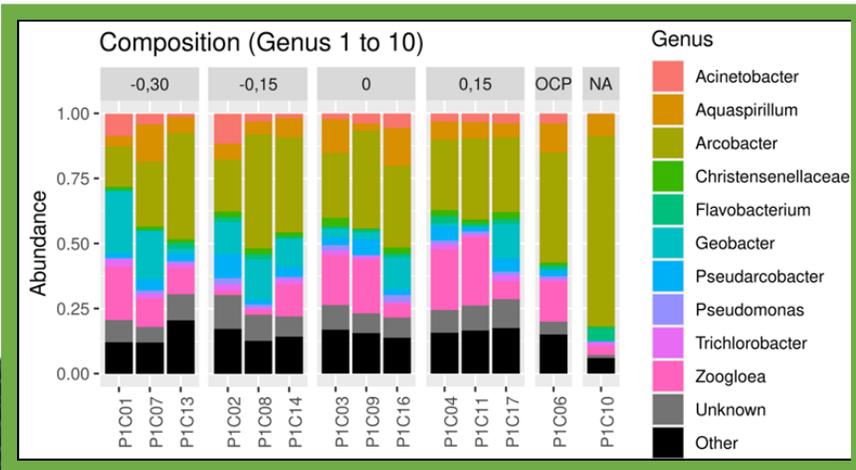
Day62

gDNA



Mainly represented in mature biofilms +150mV

cDNA



☐ Electrochemical activity

--

☐ N₂ fixation / cooperation ?

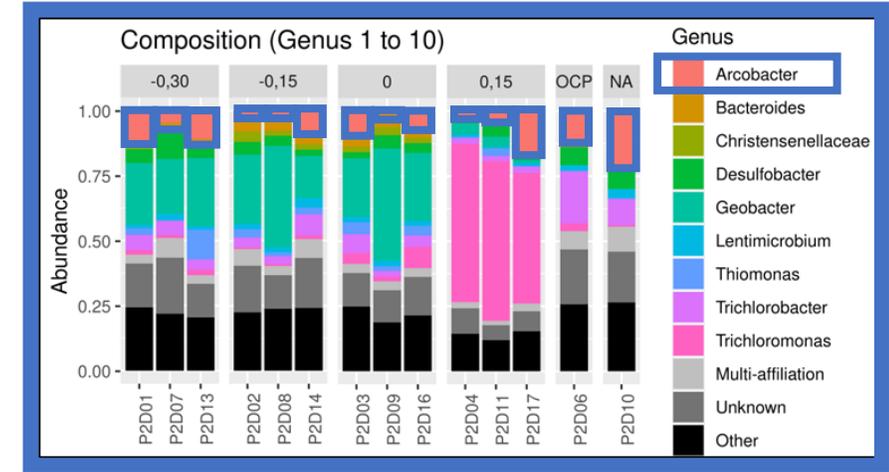
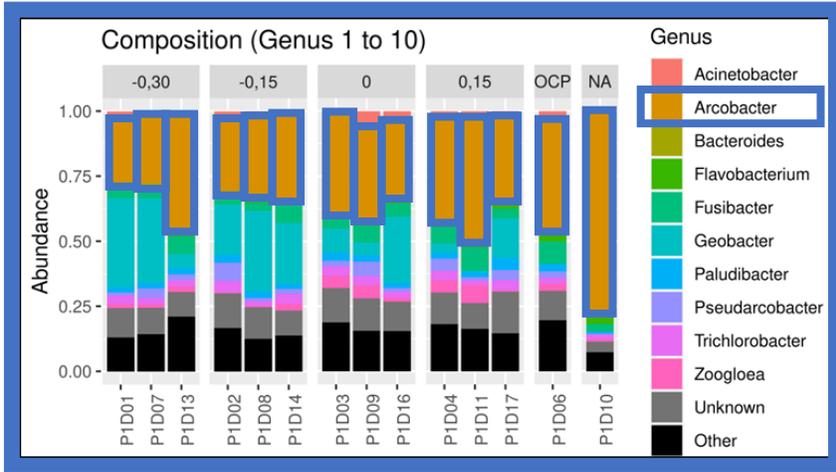


Hypothesis for *Arcobacter*

Day13

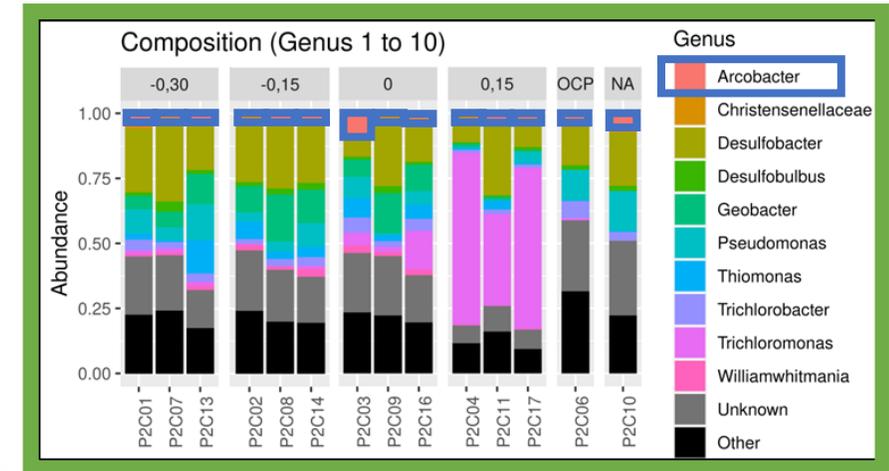
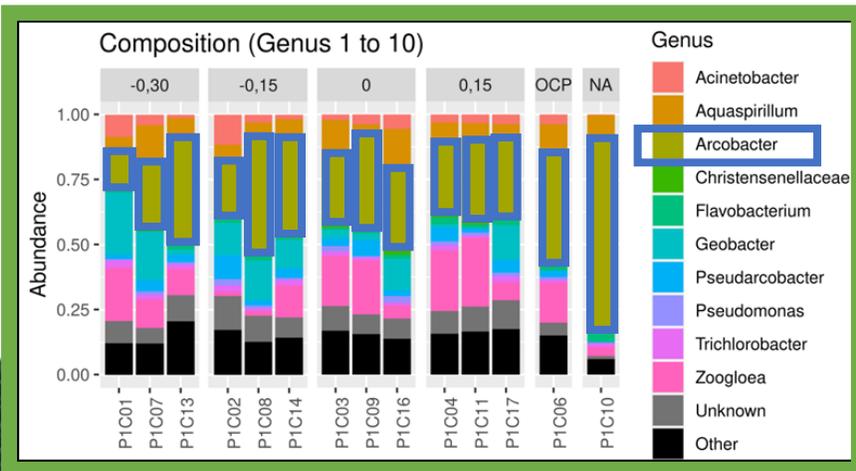
Day62

gDNA



- Mainly represented during growth phase
- Then tends to be negligible
- Within all the biofilms

cDNA



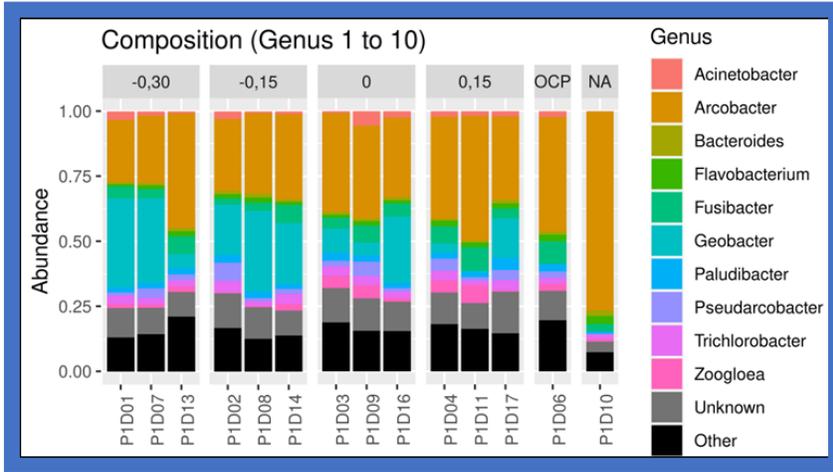
- Most important proportion
- Initial role in the biofilm adhesion?

Hypothesis for *Desulfobacter*

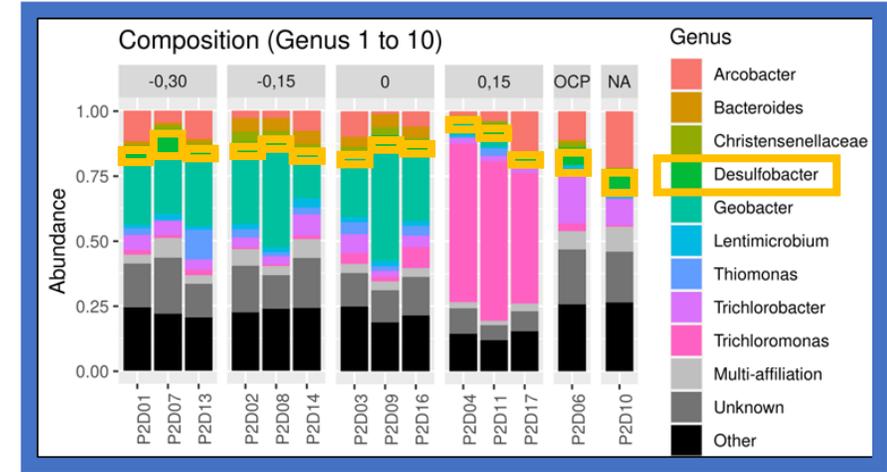
Day13

Day62

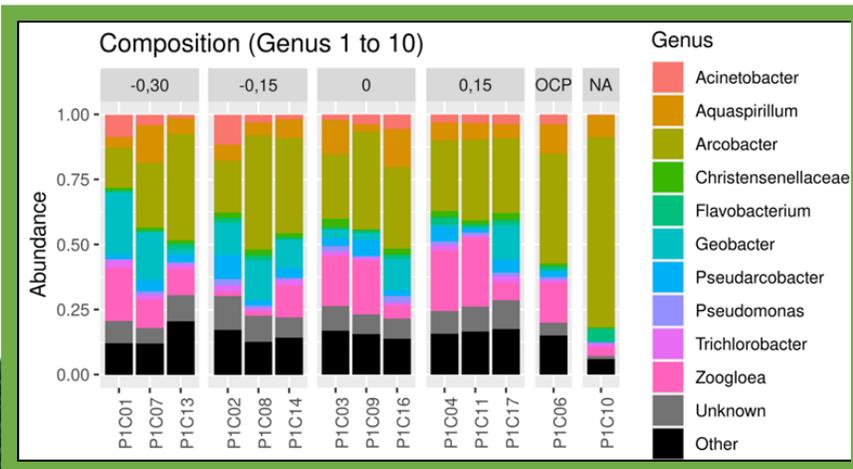
gDNA



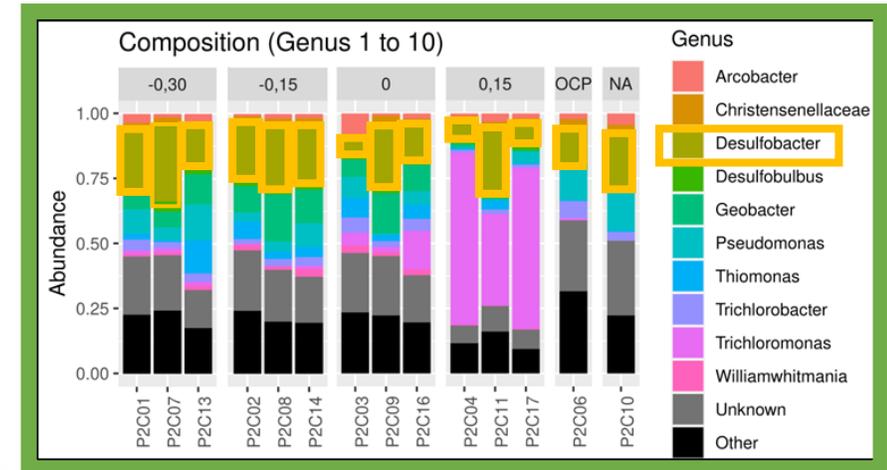
- Only present in mature biofilm
- For all imposed voltage



cDNA



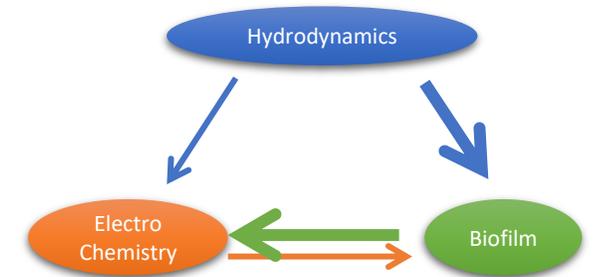
- Activity +++ in comparison to small presence
- Competition with *Geobacter* / Indicator for biofilm aging?





Take home message

- ✓ Development of a Taylor Couette electrochemical reactor validated for electro-active biofilm studies
- ✓ 3 first months of operation showing biofilm aging
- ✓ Allow to characterize electrochemically and biologically the aging





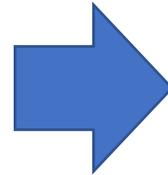
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Perspectives

- **Using the reactor to increase the shear stress:**

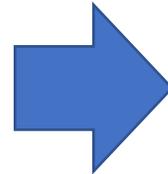
- Best accessibility to substrate?
- Detachment of *Desulfobacter*?
- More long term effect to investigate



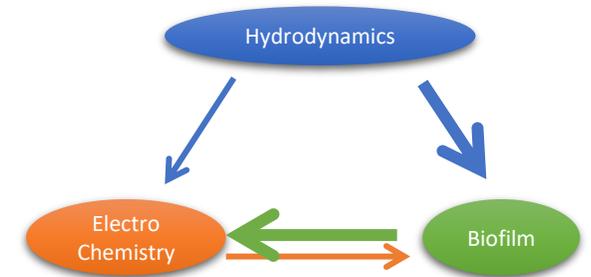
Control Tool

- **Other shear rates for biofilm growth:**

- Changes in microbial communities?
- Best efficiency or best resilience?



**Development
Parameter**





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THANK YOU.

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