

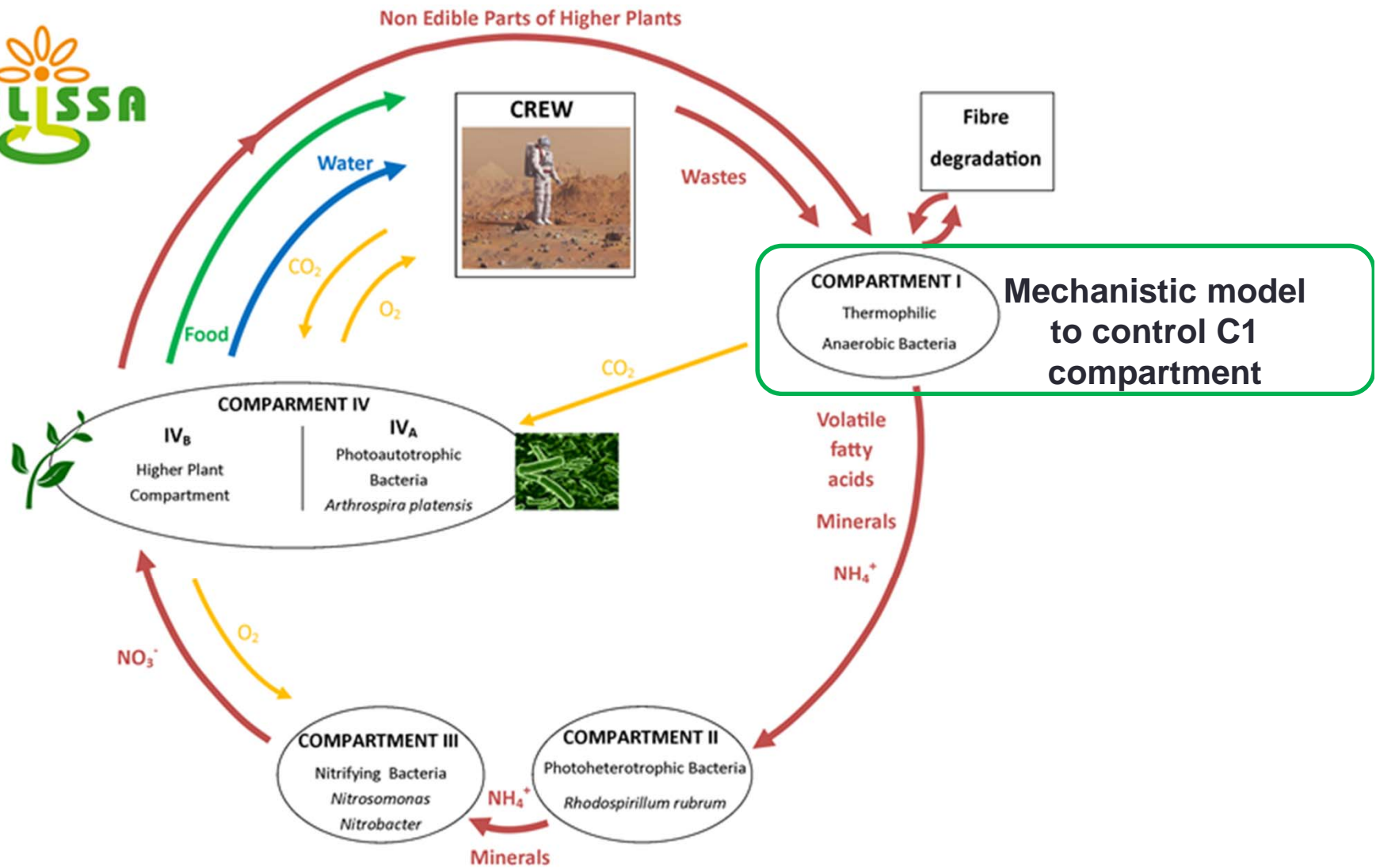
**Short and long term road map for the
development of a robust mechanistic and
dynamic model of the MELiSSA C1
compartment based on microbial community
characterization**

V. Nolla Ardèvol

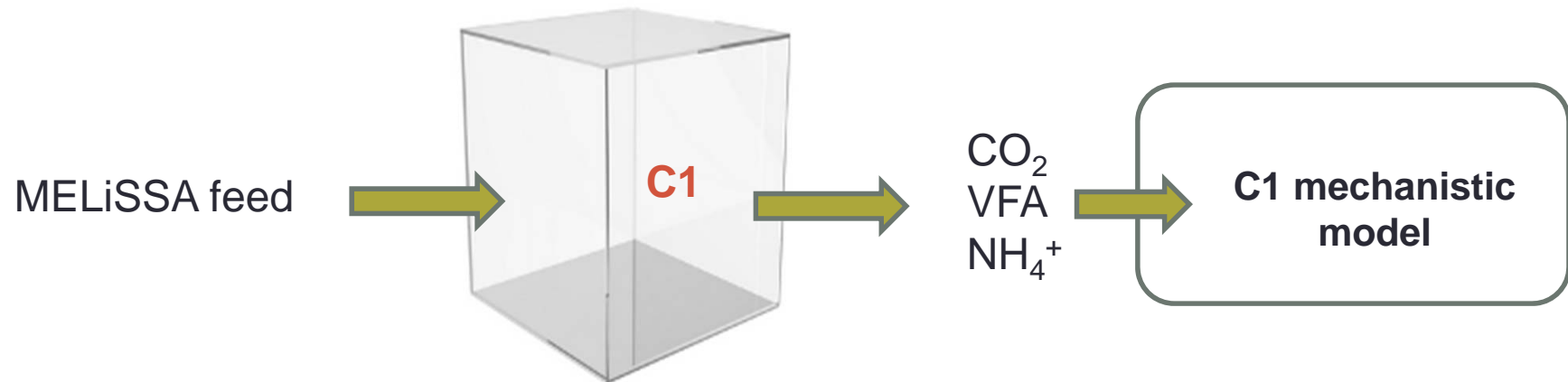
The logo for KU Leuven, consisting of the text "KU LEUVEN" in white, bold, uppercase letters on a dark blue rectangular background.

KU LEUVEN

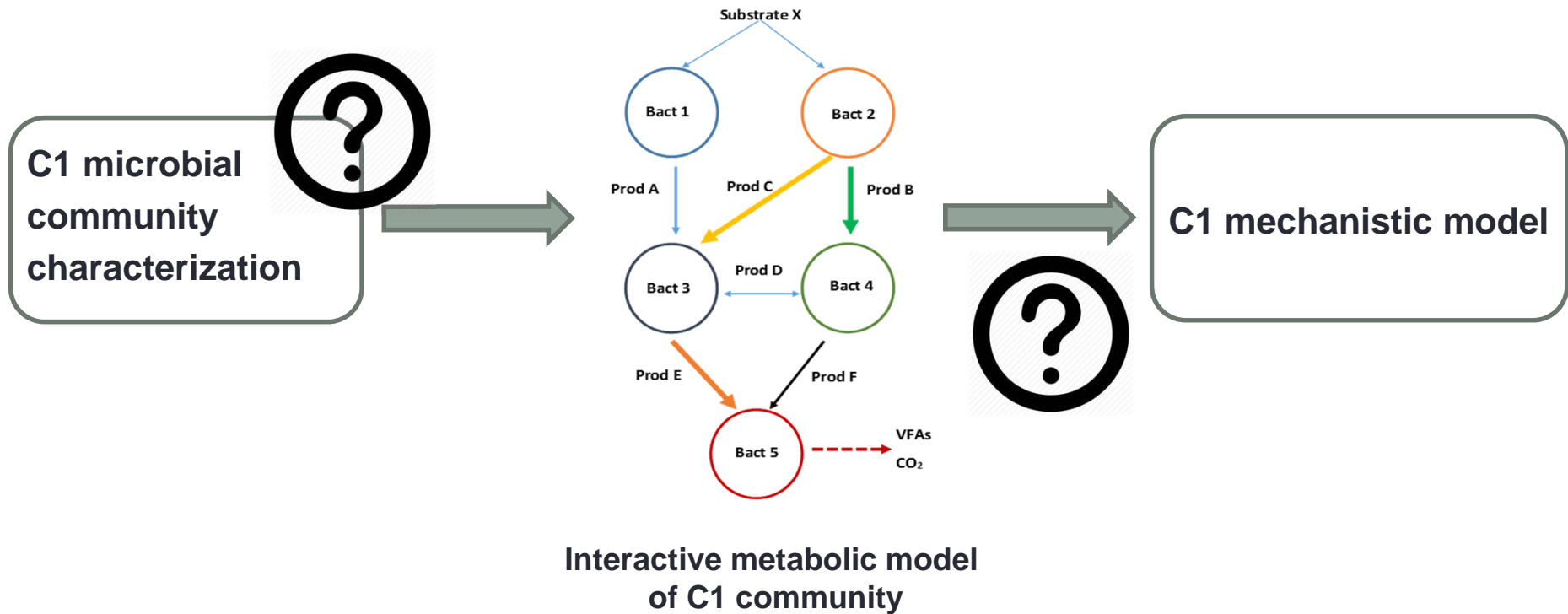
MELiSSA workshop. Lausanne.
June 8th 2016



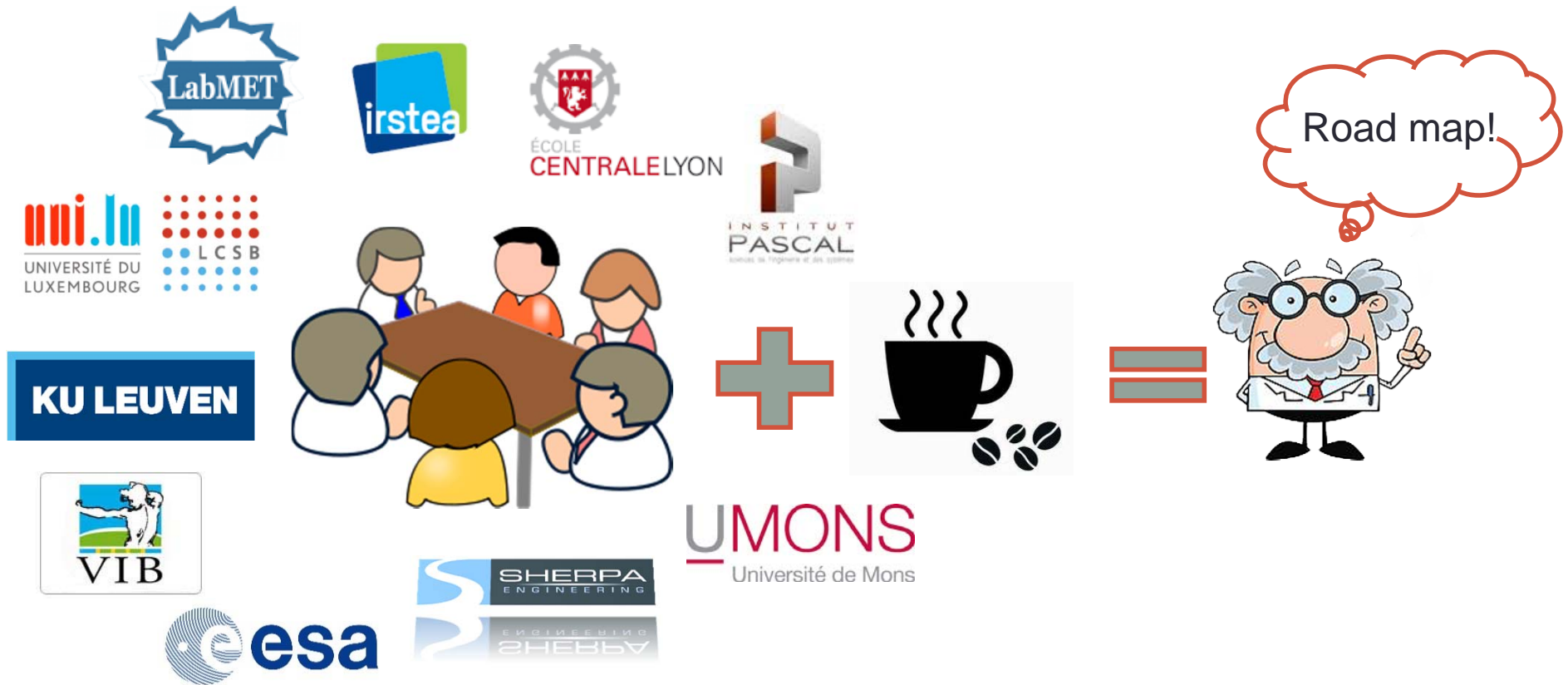
TOMORROW

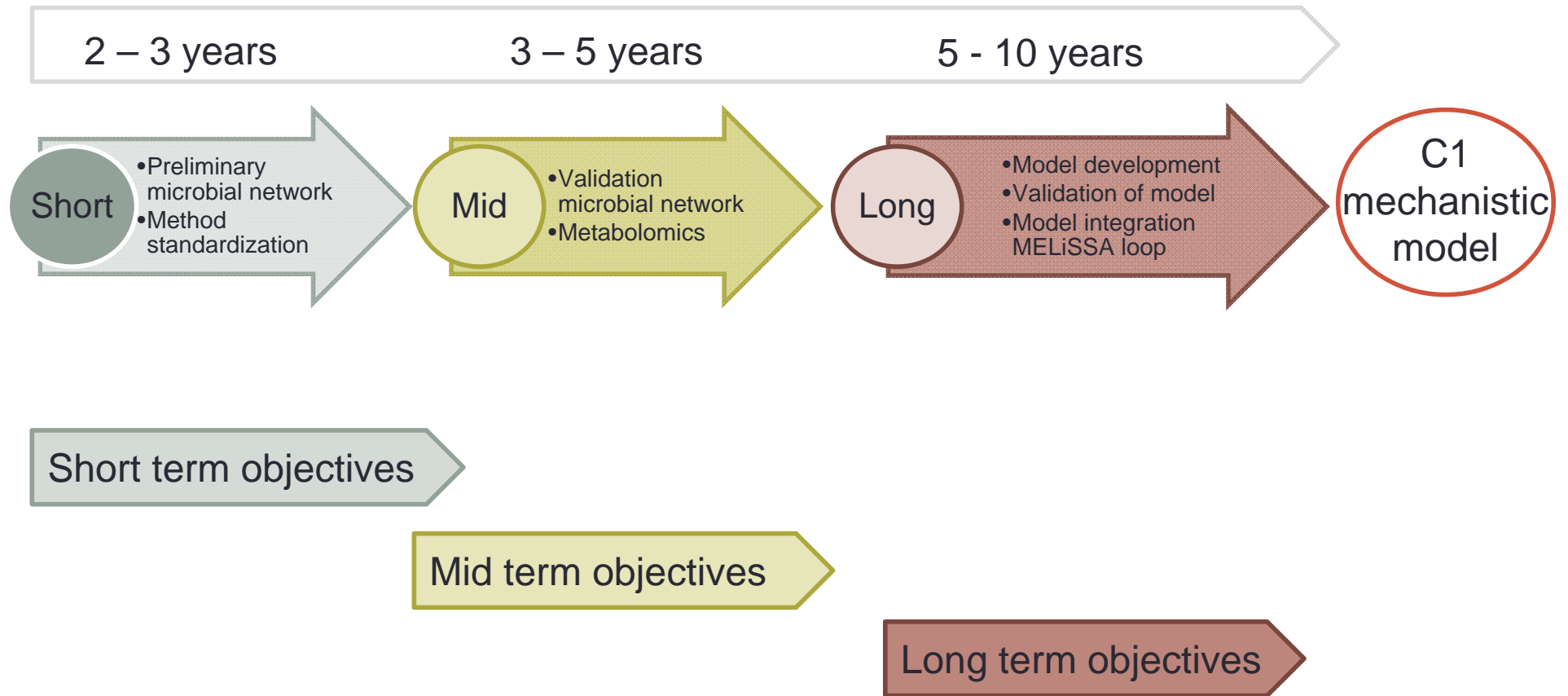


- Complete microbial information
 - Taxonomy
 - Function
 - Metabolome
- Reaction rates/kinetics
- Interactions



- **Deep Knowledge / understanding of C1 microbial community**
- **Correct experimental design / approach**
- **Which tools/techniques to use**

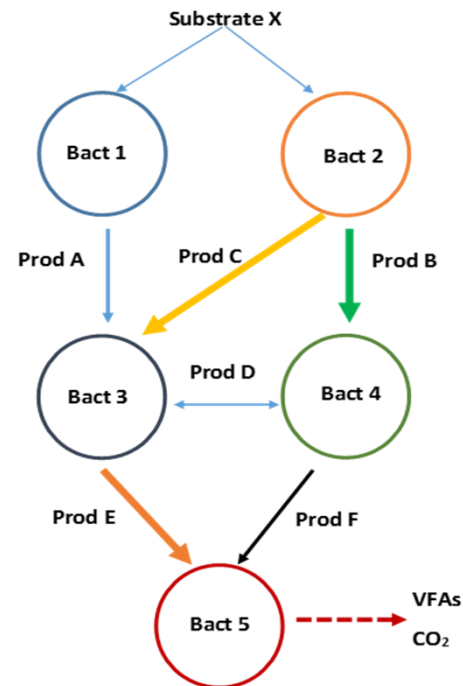




WHAT?

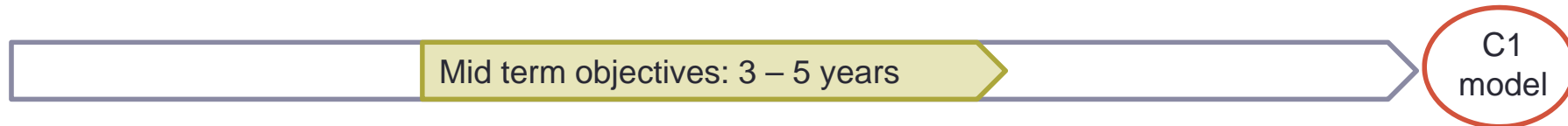
- Preliminary microbial network and microbial interactions based on a stable and well-functioning microbial community.
- Identification of crucial bacterial species and biomolecular makers involved in correct functioning of C1.

Interactive metabolic model
of C1 community
Essential functions/organisms



HOW?

- Standardization of analysis methods for omics:
 - ❖ DNA, RNA, protein extraction
 - ❖ Bioinformatic analysis
- Time point study
- Meta-omics (DNA; RNA; Proteins)
 - ❖ Determination of what is “stable/constant” (in time) community composition.
 - ❖ 16S targeted amplicon
 - ❖ Metagenomics
 - ❖ Determination of what is “stable/constant” (in time) functionality of microbial community.
 - ❖ Metatranscriptomics
 - ❖ Metaproteomics



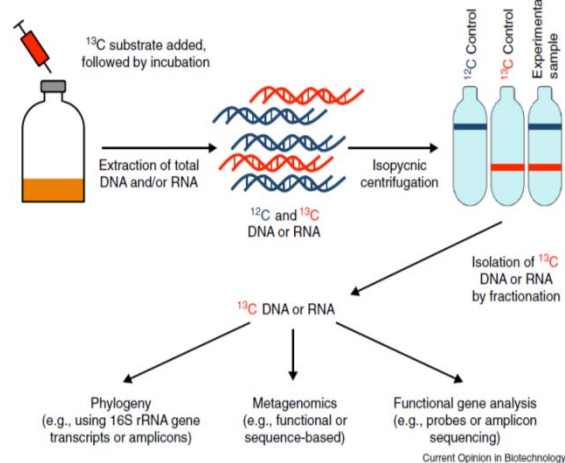
WHAT?

- Validation of microbial network proposed in short term phase.
- Study the crucial bacterial species/functions:
 - Individual species/functions
 - Selected microbial groups/functions
- Relevant perturbations to the C1 system to be able to identify differences in:
 - variations in active microbial community composition.
 - variations in the functions of the active microbial community.
- Begin with
 - Metabolomic analysis of C1 microbial community.
 - Flux analysis of selected metabolites

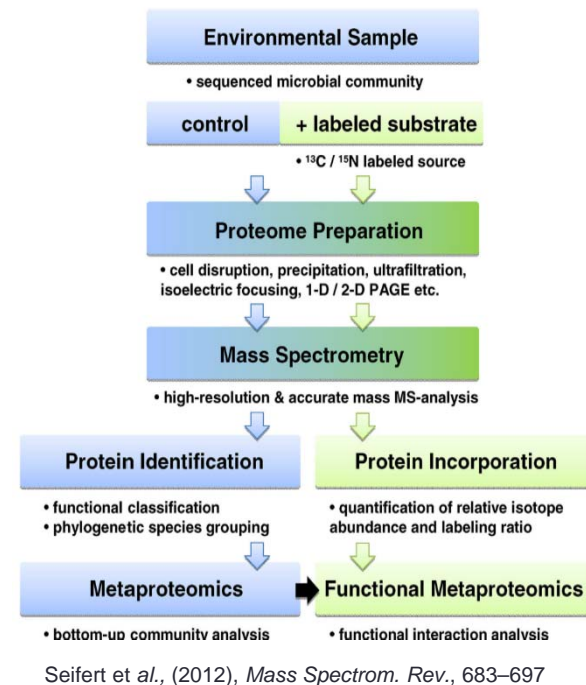
Network validation - HOW?

- SIP with different and relevant substrates to validate the microbial network.
- Identify/validate the crucial species/functions.

DNA/RNA - SIP

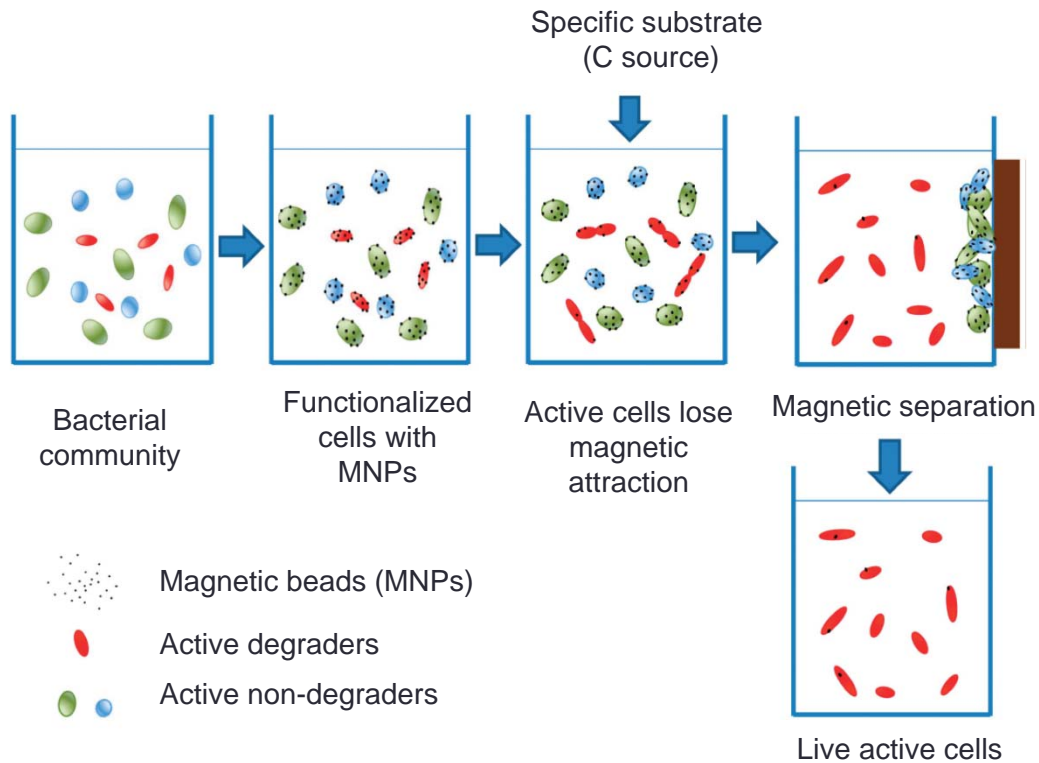


Protein - SIP

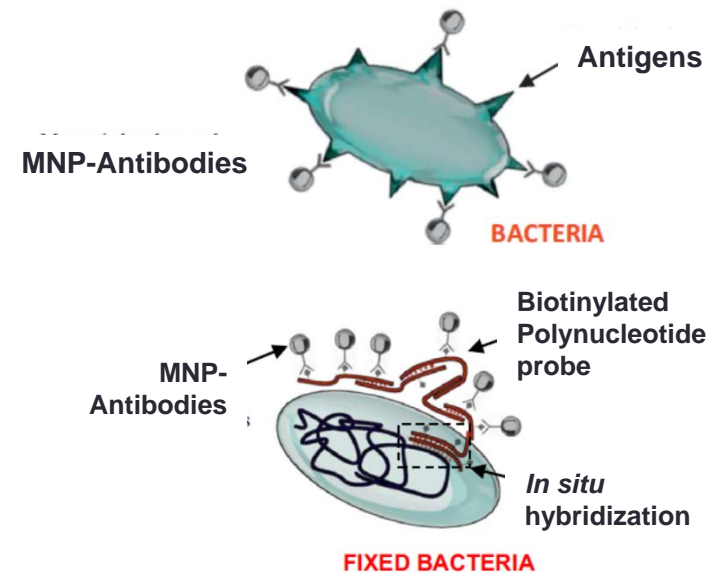


Single cells / specific populations/functions - HOW?

- Separation by magnetic nanoparticles and similar.



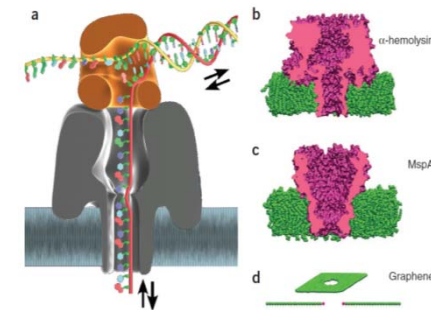
Zhang *et al.*, (2015). *ISME J.* 9, 603–14



Pivetal *et al.*, (2015). *J. Magn. Magn. Mater.* 380, 72–77.

Single cells - HOW?

- Sequencing of:
 - Single cell of relevant bacteria.
 - Metagenomic selected bacterial populations.
- New sequencing technologies:



Schneider & Dekker (2012). *Nat. Biotechnol.* 30, 326–328.



- Av read length > 10 kb
- N50 read lengths > 20 kb
- Read lengths up to 60 kb



- Av read length > 2 kb
- Read lengths up to 30 kb
- Real time results



WHAT?

- Kinetics of selected bacteria/functions.
- Kinetics of entire C1 microbial community.
- Metabolic network of the C1 microbial community.

- Integrate information for model development.

- Development technologies/assays to follow, monitor and quantify the crucial selected species, functions or biomolecular markers.

- Validation of final proposed mechanistic model.
- Integration of C1 model in entire MELiSSA loop.

HOW?



- Kinetics/ metabolomics of:
 - Selected bacterial populations/functions
 - Single cell
- Model validation with:
 - C1 perturbations/changes.
 - MELiSSA loop perturbations/changes.



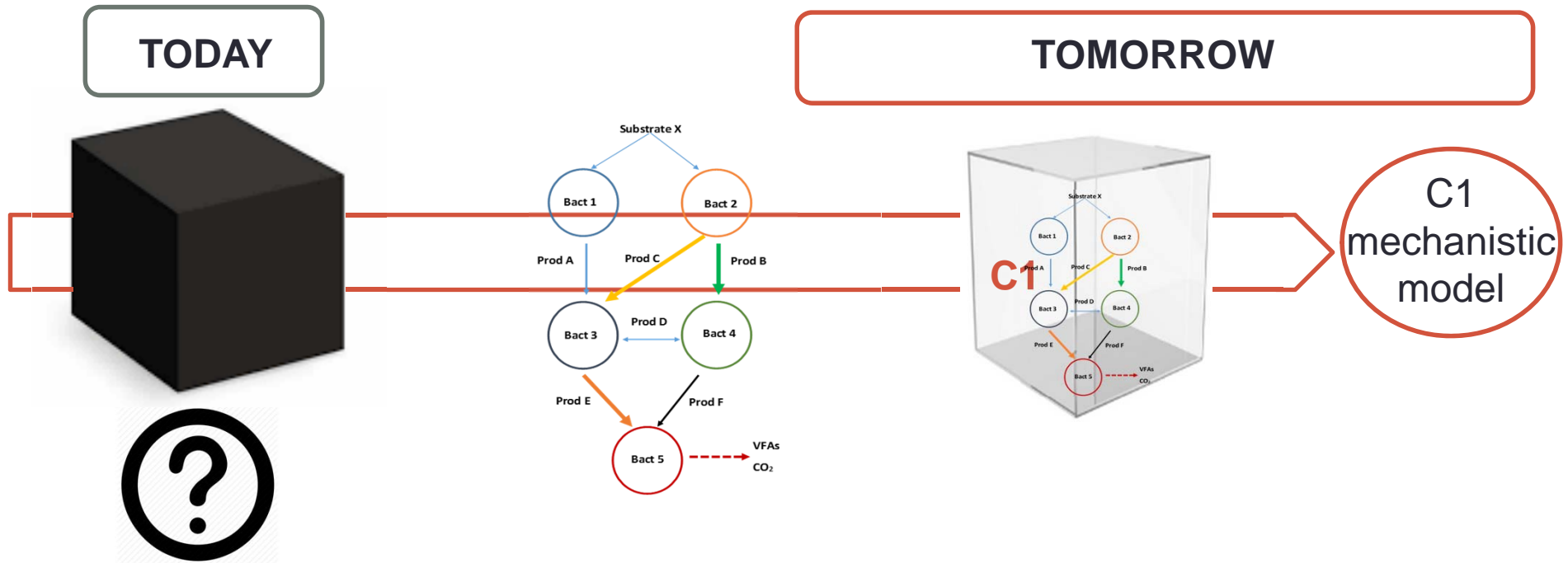
- “Miniaturized” single cell omics
- Real-time single cell/community omics
- Real-time metabolomics

Oxford **NANOPORE**
Technologies



‘SmidgION’

Summary



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