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Technology Readiness and Cultural Management Strategies for New MELiSSA Candidate Crops

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Technology Readiness and Cultural Management Strategies for New MELiSSA Candidate Crops

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1. Introduction

Advanced Life Support (ALS) systems are meant to minimize the enormous expenses associated with the resupply of life support elements during long term manned space missions. It is now accepted that the coupling of algal, bacterial and higher plant production units will more completely address the dietary needs of crew, while considering the need for stable atmospheric quality, potable water production and waste management in the manned outpost.

The European Space Agency has been involved in research relating to micro-organism based life support (MELiSSA program) and has recently contracted the University of Guelph to participate in research relating to the inclusion of a higher plant component. At present a total of eight higher crops are included in the MELiSSA program candidate list (soybean, wheat, onion, lettuce, spinach, tomato, potato, rice; Poughon, 1997). Research by NASA program members indicates, however, that a more comprehensive and nutritionally complete menu will be required (Salisbury and Clark, 1996).

A NASA workshop, entitled 'Human Nutrition in Controlled Ecological Life Support Systems' was hosted in 1996 and resulted in a preliminary list of some 45 candidate crops. These crops were selected for their yield potential, nutritional value, and psychological value and represent a consensus of scientists and engineers specializing in food production, food processing and human nutrition (JSC Memo EC3-96-130). This preliminary list was recently re-evaluated by NASA researchers in the context of the limited resources, budgets and schedules of their program. The group was instructed to focus on a narrower set of crops so that there will be a maximization of the degree to which the technological readiness level (TRL) of each crop is improved (JSC Memo EC3-98-066). As a result of these constraints, the NASA ALS program will focus on crops suitable for both vehicle and planetary food systems including dry bean, rice, sweet-potato, peanut, wheat, white potato and soybean. An evaluation of the NASA strategy in candidate crop selection has led our own researchers to the conclusion that a broader array of candidate crops should be considered regardless of their current TRL. This modified approach confers a greater ability to address dietary needs, while considering cultural compatibility, air revitalization capacity and production scheduling of biomass production units. Eleven crops including kale, carrot, peanut, sweet potato, chick pea, cow pea, lima bean, snap bean/dry bean, garden pea, lentil and broccoli have been selected for TRL evaluation and augmentation of the previous MELiSSA-HPC menu. These new crops have been evaluated through bibliographic review as was done for the eight predecessors of the MELiSSA menu.

2. Selection of Crops for TRL Evaluation

Selection of candidate crops for TRL evaluation was on the basis of four criteria, as defined by the University of Guelph research team:

- Harvest Index the ratio of dry weights of edible and inedible biomass
- •
- Nutritional Adequacy Adaptability to Closed Environment Culture Cultural Management •
- Human Factors processing, psychological appeal etc.

Much of the preliminary selection of candidate crops was based on personal experience of research team members. The task of the candidate crop evaluation was to fill in details relating to each of the four selection criteria and to assess the crops' suitability for inclusion in the MELiSSA-HPC menu. The current candidate list is presented in Table 1.

<u>Grains</u> Wheat* Rice*	<u>Starchy Roots and Tubers</u> Sweet Potato White Potato*
<u>Green Vegetables</u> Broccoli Kale	<u>Yellow Vegetables</u> Carrots
Garden Peas Lettuce*	<u>Legumes</u> Peanut
Spinach*	Soybean* Chickpea
<u>Seeds: oil & to eat</u> Peanut Soybean*	Lentil White Bean
<u>Fruits</u> Tomatoes*	<u>Condiments</u> Onion*

Table 1. Current MELiSSA-HPC candidate crops. * Indicates that crops were evaluated in TN 40.1

3. Crop Technological Readiness Level

The University of Guelph group has developed a Technological Readiness Level (TRL) scale based on the progression in research needed to achieve optimal production in a closed HPC. These TRL scales are designed to provide a metric by which candidate crops could be assessed for their readiness to be included in a complete ALS system and by which research agenda can be formulated. The TRL scales established by Eckhart (1994) for assessment and identification of flight ready hardware and the generalized crop readiness categories of NASA (JSC Memo EC-98-066) have been considered in the creation of this candidate crop TRL scale. It is important to note that progression and advancement of crops into successive TRL levels may not be linear, since time and budgetary constraints can have a profound impact on the limits of tolerance (i.e. when a crop is ready to proceed to the next research phase). It is also important to note that this TRL metric will be continuously modified as ALS engineers re-define operational and design criteria.

TRL	Type of Research	Research Focus	Planetary Food System Crop
1	Demonstration of basic production concepts	Bfield testing Byield and harvest index determinations Bnutritional value and adequacy Bplant structure considerations	Cgarden pea Cchick pea Clentil Clima bean Csnap bean/dry bean
2	Demonstration of Basic Controlled Environment Production Feasibility	Bhydroponic production possible Bidentification of adequate temperature, PPF, CO_2 , ranges for growth Bestimate yield for controlled environment	Ccow pea Ckale ÷onion
3	Quantification, modeling, and optimization of crop response	Bdose-response relationships for CO_2 and PPF, nutrients Bphotoperiod response Bphotomorphogenic responses Bgenetic and cultivar selection Byield optimization	÷lettuce ÷potato ÷rice ÷soybean ÷spinach ÷tomato ÷wheat
4	Studies of Cultural Compatibility	Bdetermine crop performance in the presence of other crops Bstabilize atmospheric conditions Bsimultaneously maintain various crop-specific conditions	
5	Integration Trials	Bcompatibility with other biomass production units and MELiSSA loop	
6	Ready for Flight validation		

Table 2. Results of Candidate Crop TRL Assessment. Crops marked with a '+' were evaluated in TN 40.1.

4. Crop Specific Cultural Requirements

The basic physiology of plant culture was discussed in TN 40.1 and the MELiSSA Annual report of 1997. Consideration has been given in the definition of plant cultural requirements to CO_2 concentration, light intensity (PPF), photoperiod, day and night temperature, and nutrient supply. For many of the newly evaluated candidate crops, specific information on cultural management strategies for closed environments could not be found in the literature and the information presented is the current state of knowledge for these crops. The information presented in Table 3 forms the basis of the assignment of TRL estimates presented in Table 2.

Сгор	CO₂ (FL/L)	Light Intensity (Fmolm ⁻² s ⁻¹)	Photoperiod (hours of daylight)	Temperature (day/night) EC	Nutrient Supply
Broccoli Brassica oleracea	—	>350	—	<9 rep.	—
Carrot <i>Daucus carota</i>	1000	>350	12-20 for vegetative	<10 rep.; 21/16 veg.	Deep Flow ½ x Hoagland's
Chick Pea Cicer arientinum	_	_	16	26/22	EC < 1.2 mmhos cm ⁻¹
Cow Pea Vigna unguiculata		>350	12	30/25	—
Lentil <i>Lens culinaris</i>	_	Ι	16	15-24 veg. 18-21 ger.	pH 5.5-6.8; EC < 3.0 mmhos cm ⁻¹
Lima Bean <i>Phaseolus lunatis</i>	_		Short Day	13-27	pH 5.5-7.0
Kale <i>Brassica oleracea</i> var. acephala	1000	>350	16	20/15	NFT ½ x Hoagland's
Garden Pea <i>Pisum sativum</i>	_	1000	16	23/16	—
Peanut <i>Arachis hypogaea</i>	_	>350	12-20	25/25 veg. 30/23 fruit	½ x Hoagland's
Snap Bean or Dry Bean/White Bean Phaseolus vulgaris	600	1000	16	24/20	—
Sweet Potato Ipomea batatas	1000	>350	<14 veg.; > 14 rep.	28/22	NFT ½ Hoagland's

Table 3. Crop Specific Cultural Management Requirements. Data presented above were compiled from a number of sources. Reported light intensities (PPF) refer to minimum recommended levels. Photoperiods listed as 12-20 indicate that the crop is photoperiod insensitive. Reported thermoperiods and photoperiods followed by veg., rep., or fruit refer to recommended periods for vegetative, reproductive (floral induction) or fruiting stages of growth. Nutrient supply is based on hydroponics delivery. NFT=Nutrient Film Technique, DFT=Deep Flow Technique. See TN 40.1 for the composition of a ½ modified Hoagland's solution. Dashes indicate that information was not available in the literature.

5. Recommendations for Further Study

Evaluation of the candidate crop TRLs indicates that many suitable species for inclusion in the MELiSSA-HPC program are not yet ready for integration into a complete ALS. Improvements in candidate crop TRL may be accomplished through an independent collection of relevant data. In particular, crops with a TRL below 3 require basic closed environment production trials in situations where key environment variables can be controlled. A series of factorial designs can be used to demonstrate feasibility while determining an initial set of cultural strategies. Crops with TRLs equal to or greater than 3 would benefit from intensive quantification and modeling studies in which CO_2 and light response curves can be generated. Such studies lend themselves to a determination of optimal cultural conditions and can be easily accomplished at the individual plant or canopy level using small sealed environment chambers.

While much of the information required for optimization of yield of the proposed candidate crops is not yet available, it is suggested that these crops comprise a portion of the MELiSSA menu. The inclusion of legumes in the menu lends itself to interesting studies relating to atmospheric nitrogen fixation. Further, it is proposed that research relating to sweet potato and peanut production be limited in the University of Guelph program since there activity by NASA and its affiliated research institutions in these areas (Morley, et al. 1999). It is also proposed that the development of dose-response models for three crops be the subject of coming research activities in the ESA-MELiSSA program. Our group is confident, then, that a good proportion of these crops can and will be included in the finalized MELiSSA menu.

6. References

General

Drysdale, A.E., Dooley, H.A., Knott, W.M., Sager, J.C., Wheeler, R.M., Stutte, G.W., Mackowiak, C.L., (1994). A More Completely Defined CELSS. Proceedings of the International Conference on Environmental Systems and 5th European Symposium on Space Environmental Control Systems, Friedrichshafen Germany, June 20-23. SAE Technical Paper Series 941292.

Drysdale, A.E., Sager, J.C., Wheeler, R.M., Fortson, R., Chetirkin, P., (1993). CELSS Engineering Parameters. 23rd International Conference on Environmental Systems, Colorado Springs, Colorado, July 12-15. SAE Technical Paper Series 932130.

Eckart, P. 1994. Life Support and Biospherics. Herbest Utz Publishers. 155

JSC Memo EC3-96-130. Baseline Crops for Advanced Life Support Program. NASA, Houston, Texas.

JSC Memo EC3-98-066. Baseline Crops for Advanced Life Support Program. NASA, Houston, Texas.

Poughon, L., (1997). Including of a Higher Plants Chamber in the MELiSSA loop: Description of a HPC for MELiSSA loop Steady State Simulations. ESA Technical Note 32.3 ECT/FG/CB/95.205.

Salisbury, F.B., Clark, M.A.Z., (1996). Suggestion for Crops Grown in Controlled Ecological Life-Support Systems, Based on Attractive Vegetarian Diets. Adv. Space Res. 18(4/5):33-39.

Broccoli

Bjorkman, T., Pearson, K.J.. 1998. High temperature arrests inflorescence development in broccoli (Brassica oleracea var. Italica). Journal of Experimental Botany 49(318):101-106

Dufault, R.J. 1988. Nitrogen and phosphorous requirements for greenhouse broccoli production. HortScience 23(3): 576-578

Fujime, Y., Hirose, T. 1984. Studies on the thermal conditions of curd formation and development in cauliflower and broccoli IV: relation between plant temperature and room temperatures in temperature controlled conditions. Technical bulletin, Faculty of Agriculture, Kagawa University, 35(2):111-120

Kojima, T., Matuzaki, K. 1995. Development of a new cultivation system by underground irrigation using porous ceramic pipes. Bulletin of the faculty of Agriculture, Saga University, 79:1-9

Kuzyakov, Y., Ruhlmann, J., Gutezeit, B., Geyer, B., Hahndel, R., Whichmann, W. 1996. Modeling on the growth and N uptake of leek and broccoli. Proceedings of the Workshop on the Ecological Aspects of Vegetable Fertilization in Integrated crop production in the field. Neustadt, An der Wein Strasse, Germany. Sept 25-29, 1995. Acta Horticulturae 428:181-191

Lougheed, E.C. 1987. Interactions of oxygen, CO₂, temperature and ethylene that may induce injuries in vegetables. HortScience 22(5):791-794

Miller, C.H., Konsler, T.R., Lamont, W.J. 1985. Cold stress influence on premature flowering of broccoli. HortScience 20(2):193-195

Miller, C.H. 1988. Diurnal temperature cycling influences flowering and node numbers of broccoli. HortScience 23(5):873-875

Mourao, I.M.G., Hadley, P., Thomas, G., Montero, A.A. 1998. Environmental control of plant growth, develpment and yield in Broccoli (Brassica oleracea var. italica Plenk): crop responses to light regime. Brassica '97. Proceedings of the International Symposium on Brassicas. Rennes, France. Sept 23-27, 1997. Acta Horticulturae, 459:71-78

Reekie, E.G., MacDougall, G., Wong, I., Hicklenton, P.R. 1998. Effect of sink size on growth response to elevated CO₂ within the genus Brassica. Canadian Journal of Botany 76(5):829-835

United Kingdom States of Jersey Dept. of Agr. 1977. Vegetables, UK States of Jersey Dept of Agriculture Report on Advisory services and Howard Davis Farm, 1975.

Carrot

Archana, M., Vora, A.B., Mancad, A. 1996. Effect of photoperiodism and GA on growth and flowering of carrot (Daucus carota L.) Indian Journal of Plant Physiology 1(4):288-9

Hebbar, R.C., Murti, G.S.R., Upready, K.K., Udaykumar, M. 1994. Effect of photoperiod, seed vernalization and growth regulators on growth, development and endogenous hormones in carrot variety Ntes. Indian Journal of Plant Physiology 37(3):137-141

Idso, S.B., Kimball, B.A., Mauney, J.R. 1988. The effects of atmospheric CO₂ enrichment on root-shoot ratios of carrot, radish, cotton and soybean. Agric. Ecos. Syst. and Environ. 21(3-4):293-299

Idso, S.B., Kimball B.A. 1989. Growth response of radish and carrot to atmospheric CO₂ enrichment. Environmental and Experimental Botany 29(2):125-139.

Krzesinski, W., Knaflewski, M. 1997. The effect of solar radiation and temperature on carrot yield and quality. Proceedings of the 5th meeting of the EUCARPIA Carrot working group, Krakow, Poland. September 1-5, 1997. Journal of Applied Genetics 38a:196-199

Mortensen, L.M. 1994. Effects of elevated CO₂ concentrations on growth and yield of eight vegetable species in a cool climate. Scientia Horticulturae 58(3):177-185 Terabayashi S. Yomo T. Namiki T. 1997. Boot development of root crops grown in deep flow and ebb and

Terabayashi, S., Yomo, T.. Namiki, T. 1997. Root development of root crops grown in deep flow and ebb and flow culture. Environment control in Biology 35(2):99-105

Wheeler, T.R., Morrison, J.I.L., Ellis, R.H., hadley, P. 1994. The effects of CO_2 , temperature, and their interaction on the growth and yield of carrot (DAU..) Plant Cell and Environment, 1994 17(12):1275-84.

Wurr, D.C.E. Hand, D.W. Edmondson, R.N., Fellows, J.R., Hannah, M.A., Cribb, D.M. 1998. Climate change: a response surface study of the effects fo CO_2 and temp on the growth of beetroot, carrots and onions. J. Agr. Sci., 131(2):125-133

Kale

Crisp, P., Crute, I.R., Sutherland, R.A., Angell, S.M., Bloorm K., Burgess, H., Gordon, P.L. 1989. The exploitation of genetic resources of Brassica oleracea in breeding resistance to clubroot (Plasmodiophora brassicae). Euphytica 42:215-226

Reekie, E.G., MacDougall, G., Wong, I., Hicklenton, P.R. 1998. Effect of sink size on growth response to elevated CO₂ within the genus Brassica. Canadian Journal of Botany 76(5):829-835

Zayed, A., Lytle, C.M., Qian, J.H., Terry, N. 1998. Chromium accumulation, translocation and chemical speciation in vegetable crops. Planta 206:293-299

Legumes

Adams M.W. and Pipoly, 1980. Biological structure, classification and distribution of economic legumes. In Advances in Legume Science, ed. Summerfield, R.J. and Bunting, A.H. Royal Botanic Gardens, Kew, England.

****Cote, R., PhD thesis

Dickson, M.H. Boettger, M.A. 1984. The effect of high and low temperatures on pollen germination and seed set in snap beans. J. Amer. Soc. Hort. Sci., 109:372-374

Duke, J.A., 1981. Handbook of Legumes of World Economic Importance. Plenum Press.

Dunlop, C.A., Ormrod, D.P. 1970. Temperature effects during fruit development on the quality of garden beans. Food Tech 3:6-8

Ellis, R.H., Summerfield, R.J., Roberts, E.H., 1993. Genetic characterization of flowering responses to photothermal environment. Aspects of Applied Biology 34:79-88

Gonzalez, R., Mepsted, R, Wellburn, A.R., Paul, N.D. 1998. Non-photosynthetic mechanisms of growth reduction in pea (Pisum..) exposed to UV radiation. Plant Cell and Environment 21:23-32

Harding, J., Tucker, C.L., and Barnes, K, 1981. Genetic variation for flowering response to photoperiod in Phaseolus lunatus L. Journal of the American Horticultural Society, 106:69-72.

Harvey, D.M. 1977. Photosythesis and translocation. In: Physiology of the Garden Pea. ed. J.F. Sutcliffe and J.S. Pate. Academic Press.

Hawtin, G.C., Singh, K.B. and Saxena, M.C., 1980. Some recent developments in the understanding and improvement of Cicer and Lens. Advances in Legume Sciences. Ed. Summerfield, R.J., Bunting, A.H., Royal Botanical Gardens, Kew, England.

Hobbs, S.L.A., Mahon, J.D. 1982. Variation, heritability and relationship to yield of physiological characters in pea. Crop science 22:773-779

Lee, Y. 1993. Temperature effects on Photosynthesis and photo-assimilate partitioning of isolated pea (Pisum sativum L.) leaf mesophyll protoplasts. M.Sc. Thesis, University of Guelph.

MacLeod, K.C., 1983. Ammonium effects on white beans (Phaseolus vulgaris cv. Seafarer) and the influence of temperature. MSc Thesis. University of Guelph.

Mahon, J.D. 1990. Photosynthetic CO_2 exchange, leaf area and growth of field grown pea. Crop science 30:1093-1098

Marsh, L.P, Davis, D.W, Li, P.H. 1985. Selection and inheritance of heat tolerance in the common bean by use of conductivity. J. Amer. Soc. Hort. Sci. 110:680-683

Ormrod, D.P. 1999. Personal communication.

Pate, J.S. 1977. The pea as a crop plant. In: Physiology of the Garden Pea. ed. J.F. Sutcliffe and J.S. Pate. Academic Press.

Siddique, K.H.M, Loss, S.P., Pritchard, D, Regan, K.L., Tennent, D., Jettner, R.L., Wilkinson, D., 1998. Adaptation of lentil (Lens culinaris Medik) to Mediterranean type environments: effect of time of sowing, yield and water use. Australian Journal of Agricultural Research 49:613-26.

Summerfield, R.J., Wein, H.C. 1980. The effect of photoperiod and air temperature on growth and yield of legumes. Advances in Legume Science, ed. R.H. Summerfield and A.H. Bunting. Royal Botanical gardens. Kew, England.

Tarila, A., 1975. The effects of phosphorus fertilization and light intensity on the growth and development of the cow pea. MSc Thesis, University of Guelph.

Wallace, D.H. 1980. Adaptation of Phaseolus to different environments. Advances in Legume Science, ed. R.H. Summerfield and A.H. Bunting. Royal Botanical gardens. Kew, England.

Peanut

Bell, M.J., Michaels, T.E., McCullough, D.E., Tollennar, M. 1994. Photosynthetic response to chilling in peanut. Crop Science 34(4):1014-1023

Leidi, E.O., Silberbush, M., Soares, M.I.N., Lips, S.H. 1992. Salinity and nitrogen nutrition studies on peanut and cotton plants. J. Plant Nutr. 15(5):591-604

Mackowiak, C.L., Wheeler, R.M., Stutte, G.W., Yorio, N.C., Ruffe, L.M. 1998. A recirculating hydroponic system for studying peanuts (Arachis hypogaea). HortScience 33(4):650-651

Nigam, S.N., Dwivedi, S.L., Ramafj, V.N., Chandra, S. 1997. The combining ability of response to photoperiod in peanut. Crop Science, 37(4):1159-1162

Sinclair, T.R., Bennett, J.M., Drake, G.M.. 1994. Cool night temperature and peanut leaf photosynthetic activity. Proceedigns og the Soil and Crop Science Society of Florida. 53:74-76

Stalker, H.T., Wynne, J.C. 1983. Photoperiodic response of peanut species. Peanut science. 10(2):59-62

Wallace, D.H., Zobel, R.W., Yourstone, K.S. 1993. A whole system reconsideration of paradigms about photoperiod and temperature control of crop yield. Theor. Appl. Genet. 66(1):17-26

Wu, W.H., Lu, J.Y., Jones, A.R., Mortley, D.G., Loretan, P.A., Bonsi, C.K., Hill, W.A. 1997. Proximate compisition and amino acid profile, fatty acid composition and mineral content of peanut seeds hydroponically growmn at elevated CO_2 levels. J. Agric. Food. Chem 45(10):3863-3866

Sweet Potato

Biswas, P.K., Hileman, D.R., Ghosh, P.P., Bhattacharya, N.C., McCrimmon, J.N. 1996. Growth and yield responses of field grown sweet potato to elevated CO₂. Crop science, 36(5):1234-1239

Hill, J, Douglas, D. David, P, Mortley, D. Trotman, A, Bonsi, C. Tozai, K. (ed.), Kubota, C. (ed.), Fujiwara, K. (ed.), Ibaraki, Y. (ed.), Sase, F. 1996. Biomass accumulation in hydroponically grown sweet potato in a controlled environment: a preliminary study. The International Symposium of Plant Production in Closed Ecosystems. Automation, Culture, and Environment. Aug 26-29, 1996. Narita, Japan. Acta. Horticulturae, 1996 440:25-30

Mortley, D., Hill, J., Lortan, P., Bonsi, C., Hill, W., Hileman, D., Terse, A., Tosai, K. (ed.). Kubota, C. (ed.) Fujiwara, K. (ed.), Ibaraki, Y. (ed.), Sase, F. 1996. Elevated CO_2 influences, yield and photosynthetic responses of hydroponically grown sweet potato. The International Symposium of Plant Production in Closed Ecosystems. Automation, Culture, and Environment. Aug 26-29, 1996. Narita, Japan. Acta. Horticulturae 440:31-36